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February 10, 2025

ELECTRONIC DELIVERY

Mr. Patrick Lennberg
Environmental Protection Specialist
Colorado Department of Natural Resources
Division of Reclamation, Mining and Safety
Office of Mined Land Reclamation
1313 Sherman Street, Room 215
Denver, Colorado 80203

Re: Permit No. M-1980-244; Cripple Creek & Victor Gold Mining Company; Cresson Project; Technical Revision 145 – Updated Grassy Valley Quality Assurance Project Plan Adequacy Review 1 Response to Comments

Dear Mr. Lennberg:

On January 28, 2025, Newmont Corporation's Cripple Creek and Victor Gold Mining Company (CC&V) received the Division of Reclamation, Mining and Safety (DRMS) Adequacy Review 1 of Technical Revision (TR) 145 to Permit M-1980-244, regarding the updated Grassy Valley Quality Assurance Project Plan (QAPP). Below are DRMS comments in bold and CC&V's response in italics.

1. **In Section 8.1 Duplicates it is stated that two duplicates will be collected during each monthly sampling event. Please update the section to clearly state from what media the duplicate samples will be collected.**

   Section 8.1 in the QAPP (Attachment 1) has been revised to specify that duplicate samples will be collected of groundwater samples.

2. **In Section 10.5 Well Sampling it is stated that wells with less than five feet of water in them will not be sampled. This is interpreted to be the case to avoid stirring up detritus from the bottom of the well which could result in analytical bias. Please provide a discussion on whether the Operator considered passive sampling techniques to sample these wells to collect an unbiased representative sample and if it would be applicable at the Site? Please provide a list of wells where there is typically less than five feet of groundwater present resulting in no samples being collected.**

   Passive sampling was not considered as an option to address this specific sampling situation, mainly due to the limited number times that this scenario arises within Grassy Valley. Upon review of the USGS's Passive Sampling of Groundwater Wells for Determination of Water Chemistry guidance document, CC&V does not believe that passive sampling would be applicable in Grassy Valley. First, it is unclear if all compliance related constituents required for analysis could be attained via a passive sampling apparatus. Secondly, the minimum required sample volume for analysis is approximately 2 liters. Assuming that the maximum volume that could be collected by a Snap Sampler would be 350 milliliters, 6 samplers would need to be deployed per well. In low-

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yielding wells with less than 5 feet of water column it is unlikely that all the samplers would fit within the water column and would not remain submerged. Finally, the potential for equipment failure, potential of degradation of the sampler, and potential damage to the monitoring well are all concerns of CC&V.

To address this issue with low-yielding wells within Grassy Valley, section 10.5 has been revised to provide a method for potential sample collection. Wells within Grassy Valley that typically have water, but less than 5-feet of water column are:

- *GVMW-24B*
- *GVMW-29*
- *GVMW-30*
- *OSABH-16*
- *OSABH-18*

Should you require further information, please do not hesitate to contact Joshua Adams at 719-851-4260 or Joshua.Adams@newmont.com or myself at Katie.Blake@Newmont.com.

Sincerely,

p.p. DocuSigned by:
Johnna Gonzalez
3AF21101E9CC41A...

Katie Blake
Sustainability & External Relations Manager
Cripple Creek & Victor Gold Mining Co

Ec: Z. Trujillo - DRMS
J. McBryde – Teller County
J. Gonzalez – CC&V
K. Blake – CC&V
N. Townley – CC&V
S. Thibeault - Newmont

KB/jg

File:Discovery\CC&V\Correspondence\DNR\DRMS\Outgoing\February 2025

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Attachment 1 – Revised QAPP

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Cripple Creek and Victor Water Monitoring Quality Assurance Project Plan and Field Sampling Guidance for Grassy Valley Monthly Monitoring

February 10, 2025

Contents

Appendices

1. Introduction

This Quality Assurance Project Plan (QAPP) for Grassy Valley monthly monitoring describes the quality assurance and quality control (QA/QC) procedures and practices for this location at Newmont's Cripple Creek and Victor Mine (CC&V). Included are standard operating procedures (SOP) for the various stages of sample collection, shipping and analyses. Sample schedules, location maps and analytical requirements are also included. This is a "living" document and is intended as guidance for site personnel; the QAPP is updated as needed to reflect changes to CC&V's water monitoring program and regulatory requirements. If a change to the QAPP is required, it will be implemented immediately and revised accordingly based on regulatory consultation as needed.

Accurate water quality data are critical to ensure permit compliance and demonstrate that water resources are not impacted by operations. The generation of reliable data begins with the collection of the sample. Adherence to the SOPs will ensure that samples are representative and collected in accordance with standard water sampling methods and QA/QC protocols. To produce data of defensible quality, this quality control program will be strictly adhered to during sample collection.

The water-sampling program includes collecting samples, recording field data, submitting samples for analyses, reviewing and recording analytical results.

1.1. Program Organization

Duties of key program personnel are listed in Table 1 below:

Table 1. Summary of Key Personnel

Name	Role	Contact Info
Katie Blake	Senior Environmental Manager	Katie.blake@Newmont.com (719) 851-4048
Josh Adams	Site Water Coordinator	Joshua.Adams@newmont.com (719) 323-0438
Trenton Reed	Environmental Technician	Trenton.Reed@newmont.com
Jordan Cranford	Environmental Technician	Jordan.Cranford@newmont.com

SVL	Contract Laboratory	TBD

- **Senior Environmental Manager:** The Senior Environmental Manager ensures the overall QA/QC program development and implementation. The Senior Environmental Manager allocates resources to ensure QA/QC and compliance criteria are met.
- **Site Water Coordinator:** The Site Water Coordinator is the program technical expert. The Site Water Coordinator oversees resource allocation, program implementation, coordinates field efforts, ensures sampling schedules are met and manages laboratory sub-contracts.
- **Environmental Technicians:** The Environmental Technicians executes the QAPP as the sampler in the field, ensuring program adherence during sample collection and shipment.
- **Contract Laboratory Manager:** Ensure analyses of environmental samples are conducted in adherence with regulatory, industry and program QA/QC requirements.

1.2. Program Objectives

The general objectives of the environmental monitoring and sampling program are:

- Collect, whenever possible, representative samples and document instances when sampling conditions are not ideal
- Ensure environmental samples are collected and analyzed according to regulatory and program QA/QC requirements
- Provide a record and insight of natural variability in environmental data as a function of seasonal meteoric changes and site evolution as influenced by historic mining practices
- Document and record environmental data history
- Identify potential environmental impacts from site activities

1.3. SAMPLING PLAN

Monitoring within Grassy Valley consists of the following:

- Monthly sampling/monitoring of 5 surface water locations
- Monthly sampling/monitoring of 34 groundwater locations

- Monthly sampling/monitoring of 5 stormwater detention ponds (i.e. EMP's)
- Monthly sampling/monitoring of seepage locations

Reporting frequencies and monitoring frequencies, locations, and parameters may be modified with the approval of the Colorado Division of Reclamation Mining and Safety (DRMS). If the monitoring or reporting requirements are modified, CC&V will update this QAPP accordingly.

2.0 Surface Water

CC&V collects surface water samples from five locations within Grassy Valley. Table 2.1 below presents the Grassy Valley Monthly monitoring compliance surface water monitoring locations. The GV-06 location is considered the point-of-compliance. Table 2.2 below depicts surface water compliance monitoring location water monitoring parameters. Table 2.3 below presents compliance limitations as specified in Colorado Regulation 32, and Figure 2.1 presents a map of the Grassy Valley monthly surface water monitoring locations.

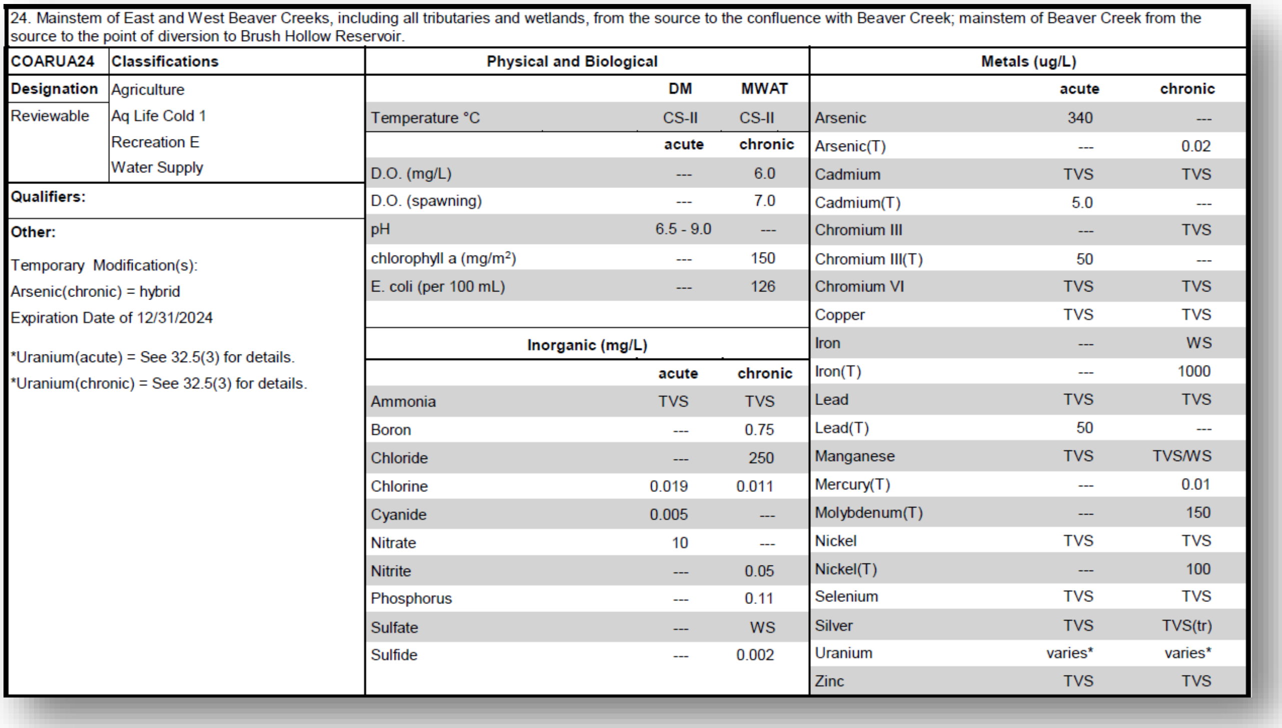
Table 2.1 Surface Water Monitoring Locations

Site Number	Location	Monitoring Frequency
GV-06	Grassy Creek - Lower Grassy Valley	Monthly
GV-4.5	Grassy Creek - Lower Grassy Valley beyond affected lands boundary	Monthly
GV-05	Grassy Creek - Lower Grassy Valley upstream of the confluence with Beaver Creek	Monthly
GV-02	Grassy Creek - Upper Grassy Valley	Monthly
GV-03	Grassy Creek - Middle Grassy Valley	Monthly

Table 2.2 Surface Water Monitoring Parameters

Parameters		
pH (Field)	Barium (mg/L) Total Recoverable	Temperature (°C)
Ammonia (mg/L as N) Total	Beryllium (mg/L) Total Recoverable	Manganese (mg/L) Total Recoverable
Cyanide [FREE] (Dissolved)	Cadmium (mg/L) Dissolved	Manganese (mg/L) Dissolved
Fluoride (mg/L)	Cadmium (mg/L) Total Recoverable	Mercury (mg/L) Total Recoverable
Nitrate (mg/L as N)	Chlorine (mg/L)	Molybdenum (mg/L) Total Recoverable
Nitrite (mg/L as N)	Chromium (mg/L) Dissolved	Nickel (mg/L) Dissolved
Boron (mg/L)	Chromium III (mg/L) Dissolved	Nickel (mg/L) Total
Chloride (mg/L)	Chromium III (mg/L) Total	Phosphorus (mg/L)
Sulfate (mg/L)	Chromium VI (mg/L) Dissolved	Selenium (mg/L) Dissolved
Aluminum (mg/L) Dissolved	Copper (mg/L) Dissolved	Silver (mg/L) Dissolved
Cyanide [WAD]	Iron (mg/L) Total Recoverable	Sulfide (mg/L)
Dissolved Oxygen (mg/L)	Iron (mg/L) Dissolved	Thallium (mg/L) Dissolved
Antimony (mg/L) Total Recoverable	Lead (mg/L) Total Recoverable	Uranium (mg/L) Dissolved
Arsenic (mg/L) Total Recoverable	Lead (mg/L) Dissolved	Zinc (mg/L) Dissolved
Arsenic (mg/L) Dissolved		

Table 2.3 Grassy Valley Surface Water Standards



24. Mainstem of East and West Beaver Creeks, including all tributaries and wetlands, from the source to the confluence with Beaver Creek; mainstem of Beaver Creek from the source to the point of diversion to Brush Hollow Reservoir.

COARUA24	**Classifications**
Designation	Agriculture
Reviewable	Aq Life Cold 1
	Recreation E
	Water Supply

Qualifiers:

Other:

Temporary Modification(s):
Arsenic(chronic) = hybrid
Expiration Date of 12/31/2024

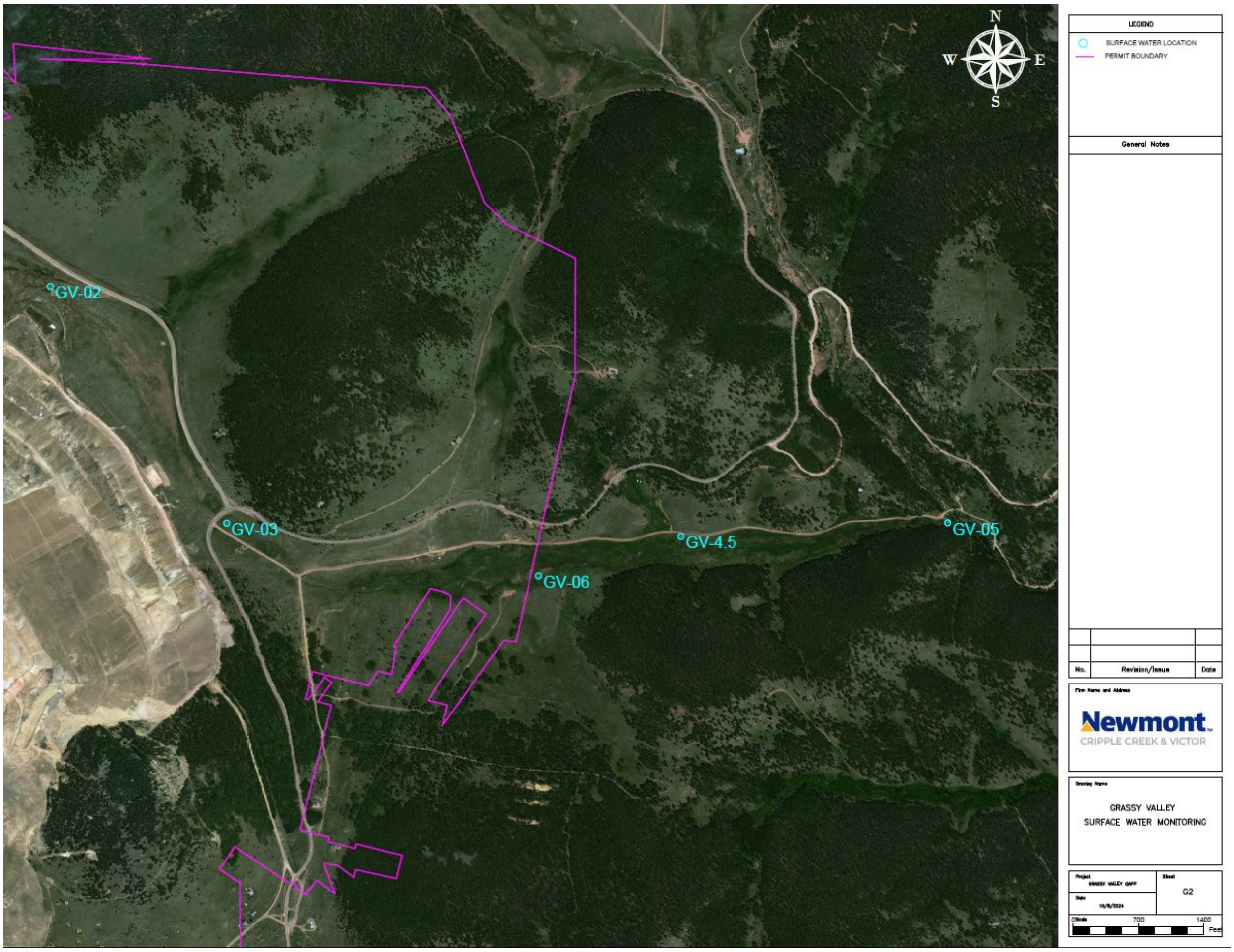
*Uranium(acute) = See 32.5(3) for details.
*Uranium(chronic) = See 32.5(3) for details.

Physical and Biological	**DM**	**MWAT**
Temperature °C	CS-II	CS-II
	acute	**chronic**
D.O. (mg/L)	---	6.0
D.O. (spawning)	---	7.0
pH	6.5 - 9.0	---
chlorophyll a (mg/m²)	---	150
E. coli (per 100 mL)	---	126

Inorganic (mg/L)	**acute**	**chronic**
Ammonia	TVS	TVS
Boron	---	0.75
Chloride	---	250
Chlorine	0.019	0.011
Cyanide	0.005	---
Nitrate	10	---
Nitrite	---	0.05
Phosphorus	---	0.11
Sulfate	---	WS
Sulfide	---	0.002

Metals (ug/L)	**acute**	**chronic**
Arsenic	340	---
Arsenic(T)	---	0.02
Cadmium	TVS	TVS
Cadmium(T)	5.0	---
Chromium III	---	TVS
Chromium III(T)	50	---
Chromium VI	TVS	TVS
Copper	TVS	TVS
Iron	---	WS
Iron(T)	---	1000
Lead	TVS	TVS
Lead(T)	50	---
Manganese	TVS	TVS/WS
Mercury(T)	---	0.01
Molybdenum(T)	---	150
Nickel	TVS	TVS
Nickel(T)	---	100
Selenium	TVS	TVS
Silver	TVS	TVS(tr)
Uranium	varies*	varies*
Zinc	TVS	TVS

Figure 2.1 Surface Water Monitoring Locations



3.0 Groundwater

Monthly groundwater monitoring locations within Grassy Valley evaluate water quality downgradient of CC&V's ECOSA facility. The GVMW-26A and GVMW-26B are considered the points-of-compliance. Table 3.1 below presents the groundwater monitoring locations for the monthly Grassy Valley monitoring, Table 3.2 presents the constituents which this location is sampled for, Table 3.3 presents the standards to which this groundwater monitoring location is evaluated against, and Figure 3.1 shows where the groundwater monitoring location is in Grassy Valley.

Table 3.1. Groundwater Monitoring Locations

Site Number	Longitude (Dec. Deg)	Latitude (Dec. Deg)	Monitoring Frequency
GVMW-4A	-105.1340	38.7505	Monthly
GVMW-7A	-105.1244	38.7477	Monthly
GVMW-7B	-105.1244	38.7477	Monthly
GVMW-8A	-105.1202	38.7413	Monthly
GVMW-8B	-105.1202	38.7413	Monthly
GVMW-10	-105.1252	38.7444	Monthly
GVMW-15A	-105.1358	38.7495	Monthly
GVMW-15B	-105.1357	38.7494	Monthly
GVMW-15C	-105.1359	38.7495	Monthly
GVMW-22A	-105.1110	38.7408	Monthly
GVMW-22B	-105.1111	38.7408	Monthly
GVMW-24A	-105.1187	38.7378	Monthly
GVMW-24B	-105.1187	38.7378	Monthly
GVMW-25	-105.1196	38.7403	Monthly
GVMW-26A	-105.1110	38.7396	Monthly
GVMW-26B	-105.1111	38.7396	Monthly
GVMW-27	-105.1217	38.7412	Monthly
GVMW-28	-105.1230	38.7422	Monthly
GVMW-29	-105.1214	38.7389	Monthly

GVMW-30	-105.1217	38.7395	Monthly
GVMW-31	-105.1221	38.7401	Monthly
GVMW-32	-105.1225	38.7406	Monthly
GVMW-33	-105.1230	38.7411	Monthly
GVMW-34	-105.1292	38.7471	Monthly
GVMW-35A	-105.1301	38.7473	Monthly
GVMW-35B	-105.1305	38.7475	Monthly
GVMW-36	-105.1337	38.7486	Monthly
GVMW-37A	-105.1181	38.7401	Monthly
GVMW-37B	-105.1181	38.7402	Monthly
OSABH-12	-105.1187	38.7377	Monthly
OSABH-14	-105.1203	38.7414	Monthly
OSABH-16	-105.1254	38.7445	Monthly
OSABH-17	-105.1328	38.7483	Monthly
OSABH-18	-105.1359	38.7495	Monthly

Table 3.2 Groundwater Monitoring Parameters

Parameters		
Aluminum (dissolved)	Cyanide [FREE]	Nitrite (NO2)
Antimony (dissolved)	Fluoride (dissolved)	pH
Arsenic (dissolved)	Iron (dissolved)	Selenium (dissolved)
Barium (dissolved)	Lead (dissolved)	Silver (dissolved)
Beryllium (dissolved)	Lithium (dissolved)	Sulfate (dissolved)
Boron (dissolved)	Manganese (dissolved)	Thallium (dissolved)
Cadmium (dissolved)	Mercury (inorganic) (dissolved)	Total Nitrate + Nitrite (NO_2+NO_3-N)
Chloride (dissolved)	Molybdenum (dissolved)	Uranium (dissolved)
Chromium (dissolved)	Nickel (dissolved)	Vanadium (dissolved)
Cobalt (dissolved)	Nitrate (NO3)	Zinc (dissolved)
Copper (dissolved)	Cyanide [WAD][1]	

[1] Weak Acid Dissociable

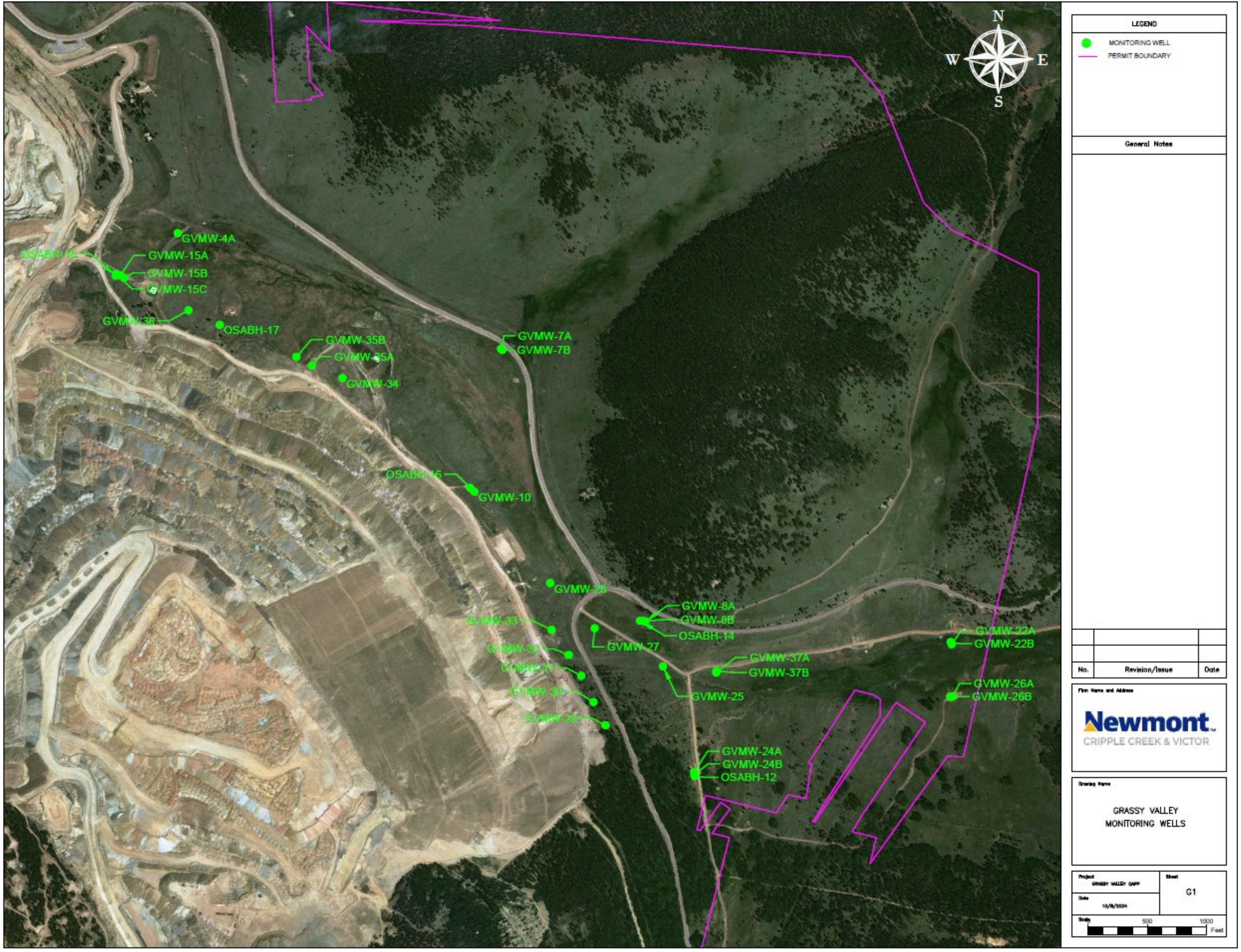
Table 3.3 Grassy Valley Groundwater Standards

Description	GVMW-8A	Sitewide NPL (Existing)	Reg. 41 Table Value Standard
Aluminum (dis) (mg/L)		7	5
Antimony (dis) (mg/L)			0.006
Arsenic (dis) (mg/L)			0.01
Barium (dis) (mg/L)			2
Beryllium (dis) (mg/L)			0.004
Boron (dis) (mg/L)			0.75
Cadmium (dis) (mg/L)		0.005	0.005
Chloride (dis) (mg/L)			250
Chromium (dis) (mg/L)			0.1
Cobalt (dis) (mg/L)			0.05
Copper (dis) (mg/L)		0.2	0.2
Cyanide [Free] (mg/L)			0.2
Cyanide [WAD] (mg/L)		0.2	---
Fluoride (mg/L)		2	2
Iron (dis) (mg/L)		14	0.3
Lead (dis) (mg/L)			0.05
Lithium (dis) (mg/L)			2.5
Manganese (dis) (mg/L)	1	3	0.05
Mercury (dis) (mg/L)		0.002	0.002
Molybdenum (dis) (mg/L)			0.21
Nickel (dis) (mg/L)		0.2	0.1
Nitrate as Nitrogen (mg/L)		10	10
Nitrite as Nitrogen (mg/L)		1	1
Nitrate + Nitrite as Nitrogen (mg/L)		11	10
pH Field - Upper	8.5	8.5	8.5
pH Field - Lower	6.5	6	6.5
Selenium (dis) (mg/L)		0.024	0.02
Silver (dis) (mg/L)			0.05
Sulfate (mg/L)	250		250
Thallium (dis) (mg/L)			0.002
Uranium (dis) (mg/L)			0.03
Vanadium (dis) (mg/L)			0.1
Zinc (dis) (mg/L)		2	2

Note:

- If no NPL exists (Sitewide or well-specific) for that parameter the most stringent value from Colorado Regulation 41 Tables 1, 2, or 3 is applied
- dis – dissolved
- mg/L – milligram per liter

Figure 3.1 Groundwater Monitoring Locations



4.0 EMPs

Enhanced Management Practices (EMPs) are stormwater collection/detention ponds that periodically have standing water. Table 4.1 below presents the EMP monitoring locations for the monthly monitoring. EMPs are sampled for the same constituents as the groundwater locations and compared to the same standards as the groundwater locations. Figure 4.1 shows the locations of the EMPs within Grassy Valley. EMPs will be sampled monthly given that they can be safely accessed, there is an adequate amount of water to be sampled, and the water is not frozen.

Table 4.1. EMP Locations

Site Number	Monitoring Frequency
EMP-16	Monthly
EMP-17	Monthly
EMP-17A	Monthly
EMP-17B	Monthly
EMP-20	Monthly

Figure 4.1 EMP Monitoring Locations

5.0 Seeps

Figure 5.1 shows the locations of the Seeps within Grassy Valley. Field parameters (pH, temperature, and conductivity) will be collected from the seep locations monthly. Samples will be collected on a quarterly basis from the seep locations and analyzed for the same parameters as the groundwater locations and compared to the same standards as the groundwater locations. Quarterly sampling of the seep locations will only occur if they can be safely accessed, there is an adequate amount of water to be sampled, and the water is not frozen.

Figure 5.1 Seep Locations

6.0 Field Technician Duties

Collection of reliable data and maintenance of analytical data are the foundation of compliance activities. Thus, the duties performed by the sampler provide the most critical element of the Environmental Department's efforts. Although this document primarily is focused on sample collection and handling methods, the generation of water data can be envisioned as a loop that includes more than simply sample collection. For any given sample, the sampler's duties have not been completed until this loop is closed. In general, a complete loop includes the collection of the sample, transmittal of the sample to a lab, receipt, review and, storage of analytical data.

6.1 Tasks

Each of these steps includes several tasks, each of which must be conducted in accordance with the procedures outlined in this document. Specific duties include the following:

- Collection of samples;
- Collection of duplicate and control samples;
- Collection of field data;
- Maintenance of equipment;
- Calibration of equipment used to collect field data;
- Tracking sample status;
- Data review and management;
- Updating sample schedules, maps and other documents as needed;
- Conducting periodic inventory of equipment;

6.2 Site Inspection

The primary function of a sampler is to collect samples. Samplers should strive to be observant of environmental conditions while in the field and should be aware of circumstances or occurrences, which are unusual or different from past events. Leaks or damp areas, materials stored in possibly unauthorized places, wildlife in the vicinity of ponds or tailing impoundments are examples of things to be noted.

Any concerns noted should be promptly brought to the attention of the area environmental coordinator.

6.3 Quality Control

Newmont's quality control program consists of the following elements: sampler competence, utilization of standards, field blanks and duplicates, calibration of meters, equipment maintenance and routine auditing of sampling procedures. Analytical results of control samples will not be used to modify any sample analyses reports.

6.4 Training

To ensure samples are collected and managed accordingly, the approved SOPs and regulatory requirements, samplers are trained by department personnel who are knowledgeable and experienced in Newmont's monitoring program. Samplers will be familiar with site history and conditions and will maintain active Mine Safety and Health Administration (MSHA) certifications. The contracted laboratory will be required to maintain appropriate certifications as needed.

6.5 Calibration

All calibration data will be documented. All field equipment will be calibrated prior to field use on daily calibration forms. Calibration procedures shall follow the manufacturers' specifications.

7.0 Monthly Grassy Sampling Report

7.1 Monthly Sampling Report

Monthly sampling reports are to be submitted to DRMS no later than 28 days after the last day of the preceding month. For example, reports are due on March 28 for February sampling event.

8.0 CC&V Monthly Grassy Sample Types

8.1 Duplicate Samples

Duplicate samples are two or more samples collected at the same time from the same location and are used to check the analyzing laboratory's accuracy. For CC&V's monthly Grassy sampling, two duplicates of groundwater samples will be collected monthly.

8.2 Rinse Blanks

A rinse blank is a sample of distilled water poured over or through a decontaminated field sampling equipment prior to the collection of environmental samples. Rinse blanks should be completed periodically to confirm that field sampling equipment is decontaminated. The field sampling technician will collect 1 rinse blank per month for laboratory analysis for the monthly Grassy sampling.

9.0 Documentation and Records

Field data is at least as important as the analytical data received from the outside laboratory. Because field data includes an evaluation of the specific instantaneous conditions at the site, this information cannot be reproduced by a later trip to the site.

9.1 Field Data

The field data is often the first indication that there may be a concern with water quality at a given location. Therefore, it is critical that any conditions observed are recorded on the field sheets.

Field data recorded at each surface, and groundwater monitoring location shall be recorded on the applicable field sheet (surface, groundwater or low flow sampling report) and will also be entered into the electronic data collection section for the Monitor Pro 5 Database Management System currently used by CC&V. Examples of the various field sheets are included in Appendix C. All field parameters collected and recorded on the completed field sheet will be entered into the electronic data entry for storage in our database. Each compliance sampling location has an associated electronic data entry form which is used to enter data.

Field data recorded at each sampling site will include, at a minimum, pH, temperature, and sampling conditions (weather, etc.). Additional data that may be recorded, depending on the specific site are dissolved oxygen, conductivity, turbidity, flow, and depth to water. In many instances, careful recording of field observations has provided clues to questionable analytical results, thus saving considerable time and money. These observations may include water color, appearance, presence of floating matter or unusual amounts of suspended material, evidence of recent activity in the area or recent access by other persons, wildlife or stock, pumping rates (for monitor well samples), or any conditions that could conceivably impact water quality. Pertinent observations during the sampling events will be recorded on the field sheets.

9.2 Calibration

All field instruments will be calibrated. The calibration and calibration check shall be documented on calibrations logs. Calibration data will not be used to alter any readings taken during the day. Calibration procedures shall follow the manufacturer's specifications.

10.0 Collection and Preservation of Samples

The objective of sampling is to collect a representative sample that ensures the analytical results accurately represent the material being sampled. Following surface and groundwater sampling SOPs ensure that this is achieved. When alternative sampling methods are necessary due to unusual circumstances, the sampler will state plainly the nature of the modification in the field sheets.

10.1 General Guidelines

A sampling event (day) should be scheduled to collect background samples first, and samples such as process solutions last, to avoid cross contamination of wells or streams. In general sampling should begin with locations that have the lowest concentrations of constituents and move to higher concentrations throughout the day.

Samples will be collected in new sample bottles of material consistent with the parameters to be analyzed. DO NOT touch the inside of sample vessel or cap or allow these surfaces to contact any material other than the sample media. Sample containers that are known, or suspected, to be contaminated will be discarded or clearly marked with an "X" or other designation to prevent their use. Holding times, minimum required sample bottles/volume, and necessary preservative types are contained within attachment I.

Table 10.1.1 below contains the required sample bottle, bottle volume, and preservative for the various analysis suites CC&V uses for our water monitoring program.

Table 10.1.1 - Sample Bottle Requirements by Analysis Suite

Surface Water Analysis Suite
1 - 500 mL HDPE unpreserved sample (grab sample) Black Label
1 - 250 mL HDPE unfiltered sample (grab sample) preserved with 1.25 mL H2SO4 (Sulfuric Acid, yellow label)
1 - 250 mL HDPE unfiltered sample (grab sample) preserved with 1.25 mL HNO3 (Nitric Acid, red label)
1 - 250 mL HDPE filtered sample (grab sample) preserved with 1.25 mL HNO3 (Nitric Acid, red label)
1 - 250 mL Amber HDPE unfiltered sample (grab sample) preserved with 1.25 mL NaOH (Pre-preserved bottle or Sodium Hydroxide, green label) 1 - Hexavalent Chromium Kit (pre-preserved) adjust pH as needed with NaOH (Sodium Hydroxide, Green Label) to achieve pH between 9.3 - 9.7. (All preservatives included in kit, along with instructions and pH strips). 1 - 250 mL HDPE filtered sample (grab sample) preserved with 1.25 mL Zn Acetate NaOH (Zinc Acetate NaOH, purple label)
Groundwater Analysis Suite
1 - 500 mL HDPE unpreserved sample (grab sample) Black Label
1 - 250 mL HDPE unfiltered sample (grab sample) preserved with 1.25 mL H2SO4 (Sulfuric Acid, yellow label)
1 - 250 mL HDPE unfiltered sample (grab sample) preserved with 1.25 mL HNO3 (Nitric Acid, red label)
1 - 250 mL Amber HDPE unpreserved sample (grab sample) preserved with 1.25 mL NaOH (Sodium Hydroxide, green label)
1 - 250 mL HDPE filtered sample (grab sample) preserved with 1.25 mL HNO3 (Nitric Acid, red label)

Disposable latex surgical gloves will be worn during the collection and preservation of samples to minimize potential contamination of the sample, and to protect hands from preservatives and process water.

When filling containers leave a small air space to allow for thermal expansion unless sampling for organics or dissolved oxygen which requires zero head space.

10.2 Sample Field Sheets

A detailed record will be made at the time of collection of all pertinent information related to the sample. See Field Data in the previous section (Section 9.0) for appropriate information. For locations requiring low-flow sampling protocols, a low flow sampling report will be generated to document the process. Low Flow Sampling Reports are included in Appendix C and the procedure is described in Appendix I.

10.3 Sample Identification

Gummed paper labels or tags will be filled out with waterproof ink at the time the sample is collected. The labels should contain the following information: date and time of sample collection, sample location, sample identification (ID#), name of sample collector, whether the sample was filtered, and type of preservative used. The labels must be attached to the appropriate sample bottle. In the absence of labels, write the above information directly on the sample bottle with a permanent marker.

Care must be exercised to ensure that the sample ID# is the same as the official designation for each sample location. Failure to use the same ID# as specified in the applicable permit may result in analytical results being questioned. Officially designated sample ID's for compliance sampling locations are specified in appendix G. Non-compliance samples to be collected shall be collected as necessary, with all necessary information being recorded within the sampler's field notebook. It is extremely important that sample identification and recorded notes be sufficient to identify precisely where the samples came from.

Duplicate and control samples will be identified with an ID# which will not bias the laboratory by indicating the origin of the sample. At CC&V duplicate samples are identified by increasing the numerical component of the monitor well identifier by 100, and offsetting the position nomenclature by positive 5 (Increasing the letter count by 5). For example the duplicate for monitoring well VIN-2A would be VIN-102F, the duplicate for monitoring well GVMW-22B would be GVMW-122G. It is also necessary that all pertinent field data for the duplicate sample be recorded as any sample, and that the sample ID is identified as a duplicate, and which well it is a duplicate for is identified.

Blank samples will be identified with an ID# which will not bias the laboratory by indicating the origin of the sample. At CC&V Blank samples are identified by using the prefix CCVB and then following it by the month numerical value and the day numerical value (two digit). For example the blank sample collected on March 25 would be identified as CCVB-0325. It is also necessary that field data be collected for the blank sample (pH, DO, EC, temp). The field data needs to be recorded on a field sample sheet.

10.4 Sample Collection

Decontaminate all non-dedicated sampling equipment and meters, as appropriate, before and after use with deionized water (Type III reagent grade). Decontamination procedures include cleaning of equipment with a dilute phosphate free detergent solution (i.e. Alconox or Liquinox), followed by a fresh water rinse.

To assure an undiluted sample is collected, field cups, filter vessels, or other reusable equipment should be triple rinsed with sample solution if sufficient quantities are available.

Field readings will be measured from a separate container collected at the same time as the sample, and will not be taken from the actual sample bottle which will be analyzed. If a field sample was taken from a lined facility, it must be returned to a lined facility. Minimum field readings will be pH, conductivity, and temperature.

Sample collection from well, stream, pond, reservoir, & waste rock discussed below.

10.5 Well Sampling

To begin each sampling event, measure depth, to the nearest one-tenth of a foot, to static water level from the top of casing (TOC) with a water level indicator (Solinst). Rinse the level indicator with deionized water (type III reagent grade) before and after use. A dry well will be recorded as "Dry at X feet" to assure that the Solinst did not hang up in the well.

Water standing in a well prior to sampling is not representative of in-situ ground water quality. Therefore, the stagnant water must be removed and replaced by fresh formation water. EPA protocol dictates that one to ten volumes of water standing in the well casing should be removed prior to collection of the sample. At CC&V the standard well sample collection method is the EPA Low Flow methodology (Low-Flow (minimal drawdown) Ground-water sampling procedures).

When sampling using EPA Low-Flow methodology, start the pump at low speed and slowly increase the speed until the discharge occurs. Record the pumping rate for future sampling and try to match the pumping rate used during previous sampling events. Otherwise adjust the speed until there is little to no water level drawdown. If the minimal drawdown exceeds 0.3 feet, but remains stable, continue purging. Monitor and record the water level and pumping rate at least every five minutes during purging. Record any pumping rate adjustments (both time and flow rate). Pumping rates should, as needed, be reduced to the minimum capabilities of the pump to ensure stabilization of the water level. Adjustments are best made in the first fifteen minutes of pumping in order to help minimize purging time. During pump start-up, drawdown may exceed the 0.3 feet target and then "recover" somewhat as pump flow adjustments are made. Purge volume calculations should utilize stabilized drawdown value, not the initial drawdown. If the initial water level is above the top of the screen do not allow the water level to fall into the well screen. The final purge volume must be greater than the stabilized drawdown volume plus the pump's tubing volume. If the drawdown has exceeded 0.3 feet and stabilizes, calculate the volume of water between the initial water level and the stabilized water level. Add the volume of the water which occupies the pump's tubing to this calculation. This combined volume of water needs to be purged from the well after the water level has stabilized before samples are collected. During purging monitor field parameters (pH, temperature, & conductivity). Purging is considered complete and sampling may begin when all the indicator field parameters (below) have stabilized. Achievement of turbidity levels of less than 5 NTU, and stable drawdowns of less than 0.3 feet, while desirable, are not mandatory.

The EPA sampling method is designed for ideal conditions and wells. Wells and conditions at CC&V can be variable and it should always be the goal of this sampling program to collect meaningful data. If deviations are made from the standard procedure, they will be documented and done so in the interest of collecting data that is useful.

Some wells yield such low volumes of water that this protocol cannot be followed. Wells containing less than 5 feet of water column are considered low-volume or low-yielding wells. When sampling a low yield well evacuate the well to dryness once. Within 24 hours of this purge, if the well has recovered at least 90% of the original volume collect, preserve, and handle the sample(s) according to normal procedures. This procedure described mimics that described by the United States Geologic Survey's National Field Manual for the Collection of Water-Quality Data.

If a low-volume well (less than 5' of water column) does not purge to dryness after 3 casing volumes are removed, follow the standard/well volume method.

Standard/Well-Volume Method may also be utilized for sampling monitoring wells, for this method three casing volumes will be evacuated prior to sampling. Measure the stabilization criteria outlined below after each well volume is evacuated (i.e., if the well volume is 5 gallons, take measurements after evacuation of 5, 10, and 15 gallons). On the field form, record the volume of water evacuated, all stabilization parameters, and time that the measurements were made. After three well volumes have been purged (appendix E, well evacuation calculation) check the last two sets of measurements to determine if the field parameters have stabilized. If the field readings have not stabilized purge another well volume and take field measurements. Repeat until stabilized.

Stabilization criteria are achieved when the following parameters are met over three consecutive readings.

- **Temperature** - ±3%,
- **Specific Conductivity** - ±3%
- **pH** - ± 0.1
- **Dissolved Oxygen** - (10% for values greater than 0.5 mg/L, if three Dissolved Oxygen values are less than 0.5 mg/L, consider the values as stabilized)
- **Turbidity** - (10% for values greater than 5 NTU; if three Turbidity values are less than 5 NTU, consider the values as stabilized)

- **Oxidation/Reduction Potential** - ±10 millivolts

If the field values indicate stable conditions, collect, preserve, and handle the samples according to the procedures outlined in this document. An example of a well purging calculation is included in Appendix E.

10.6 Stream Sampling

Sampling results will vary with depth, stream flow, and distance from shore. When rinsing the field cup or bottles, discard the rinsate downstream from the sampling point. A grab sample should be collected at mid-depth from the middle of the stream, in an actively flowing section of the stream. In shallow streams, care should be taken not to disturb the bottom and put sediments into suspension, as these will affect the analytical results. If a stream has no visible flow, it will be recorded as dry or stagnant and will not be sampled. If a stream sampling location has visible water, but not enough water to allow collection of a representative sample, it will be recorded as "too low to sample". Sampling data will be recorded on the sampling logs.

10.7 Filtering Samples

A ground water sample to be analyzed for dissolved metals must be passed through a 0.45 micron membrane filter prior to preservation. For the determination of total metals, the sample is not filtered. As a general guideline, ground water (wells) samples requiring a metals analysis should be filtered and analyzed for dissolved metals, while surface water samples requiring a metals analysis should be unfiltered and analyzed for total metals. Samples requiring analysis for organics should not be filtered. Specify on the Newmont chain of custody whether or not the samples have been filtered.

10.8 Sample Preservation and Storage

Sample preservation is intended to retard breakdown of the constituents within the sample. Preservation methods include pH control, chemical control, temperature control, and protection from light. Common sample preservation measures include the following:

To avoid changes in the concentration or physical state of the constituent to be analyzed, preserve accordingly, and pack samples in ice in the field and when shipping. Samples should be stored in a locked refrigerator and shipped to the laboratory as soon as possible. As samples are transferred to the storage refrigerator, a double check that the lids are securely tightened is conducted.

10.9 Sample Handling and Custody

The contracted laboratory will provide necessary coolers, sample bottles, chain-of-custody (COC) forms and shipping labels. After sample collection, samples will be stored in a refrigerator and shipped to the laboratory as soon as possible and within allowable holding times. Samples will be cooled to temperature acceptable to the contracted labratory. Sample containers will be packed to prevent breakage or contamination during shipment.

10.10 Chain of Custody Procedures

Chain of custody procedures will allow for the tracking of individual samples from the time of collection through laboratory analysis. All records relating to chain of custody documentation are to be made in ink. If errors are made on any of these documents, corrections are to be made by crossing a single line through the mistake and entering the correct information. All corrections are to be initialed and dated by the individual making the error, if possible, or by the investigator. All paperwork completed in the course of collecting and shipping samples must be correct, accurate, and defensible in a court of law.

The complete COC will accompany the sample from the site, through delivery to the contracted laboratory. To ensure complete documentation of sample custody, field personnel and laboratory personnel will sign and date the COC upon shipment and receipt.

10.11 Sampling Field Sheets

The sampling logs contains the first record in the chain of custody of the sample. It is previously discussed in Section 9.0 (Field Data) of this document.

10.12 Chain of Custody Record

A chain of custody record will be completed and must accompany each sample or each cooler of samples (see Appendix A). The record will include the following information: specific area/permit name, sample identification, sample type (well, grab, soil, other), preservative(s) used, whether the sample was filtered, type of analysis(es) required, number of sample containers, signatures of persons involved in the chain of possession, inclusive dates of possession, a unique sample ID, date, and method of shipment. If an analysis is to be rushed, state this on the Chain of Custody. One copy of the chain of custody is kept by Newmont. The laboratory keeps one copy for its records, and returns a copy to Newmont with the analysis reports.

10.13 Shipping Papers

A shipping label is attached to the top of cooler or shipping container along with a laboratory's address label. The cooler or shipping container must be secured with shipping fasteners or packing tape to prevent opening during transportation.

10.14 Delivery to Laboratory

Planning is required to ship the samples, so the lab personnel are available to receive them, especially if shipping over the weekend. If a rush analysis is requested, notify the laboratory ahead of time. To ship samples, fill out a shipping paper for each cooler (see Appendix C) and deliver the coolers and shipping papers to the shipper. Make a copy of the shipping paper, COC and file in the Chain of Custody folder (see Tracking System section below). **Note**: For water quality samples there must be enough ice in the cooler to ensure the sample temperature is maintained at a temperature that is acceptable to the laboratory.

10.15 Data Receipt and Review

Analysis reports are received with the invoices. The data should be reviewed relative to the applicable standards. Should any analytical results deviate from expectation or exceed the applicable standards the coordinator should be informed.

11.0 Well Sampling Equipment

If possible, dedicated sampling devices should be used in water quality monitoring. When non-dedicated equipment is used, equipment will decontaminated using the procedures detailed above, before and after use to avoid cross contamination of wells. Equipment should be used according to manufacturers' guidelines. A brief summary of guidelines for various types of equipment used by CC&V follows:

11.1 Dedicated Electric Powered Sampling Pump

When possible, use the dedicated electric powered sampling pump to purge sampling wells. If the electrical connection coming from the dedicated sampling pump is a 240 volt connection, connect directly to the 240 volt input on the generator. If a four prong electrical connection comes from the dedicated sampling pump, connect the four prong connection from the sampling pump to the four prong connection on the control box. Then connect the 240 volt connection on the flow control box with the 240 volt input on the generator. Once the electrical connections are made pull out the choke on the generator, ensure the gas valve is open and start the generator. Once the generator is started push in the choke, if the control box is connected, adjust the flow rate using the nob on the control box. The optimum flow rate should allow at least three well volumes to be purged without purging the well dry.

After sampling turn off the generator and close the gas valve. Disconnect the electrical connections from the sample pump, control box, and generator. Ensure the sampling pump's electric connection and wiring is inside the well casing and secured below the well cap.

11.2 Field Deployable Submersible Pump

When a dedicated electric powered sampling pump is not available for use at the monitoring well to be sampled, a field deployable submersible pump is to be used.

Currently CC&V uses a Geotech Environmetnal SS Geosub or a Grunfos Redi-Flo 2 portable submersible pump for sample collection. This submersible pump has an external controller to manage the pump. To use the pump follow the instructions and training provided to you, if you have not received the necessary or adequate instruction contact your supervisor to receive the necessary training. Insure that all electrical connections are solid, and that the necessary grounding is in place. Operate the necessary power equipment (generator or inverter) to power the controller to collect the sample.

After sampling turn off the generator and close the gas valve (if used). Disconnect the electrical connections from the sample pump, control box, and generator.

11.3 Bailer

The hand bailer is a cylindrical tube, constructed of either plastic or metal, with a ball valve at the bottom. The ball valve allows water into the tube while the bailer is descending and closes, thus trapping water, when the bailer is ascending.

The bailer is to be decontaminated as described above, before and after use. The bailer is lowered into the well with a nylon rope, or a stainless-steel mesh rope, which is decontaminated after each use. Care must be taken to ensure that the rope does not come into contact with the ground while sampling, to avoid any possible contamination. A disposable bailer must be used and then discarded when sampling for VOCs. Otherwise, it may be decontaminated and reused.

The line should be securely affixed to the bailer, and the opposite end should be securely affixed to something to prevent losing the bailer

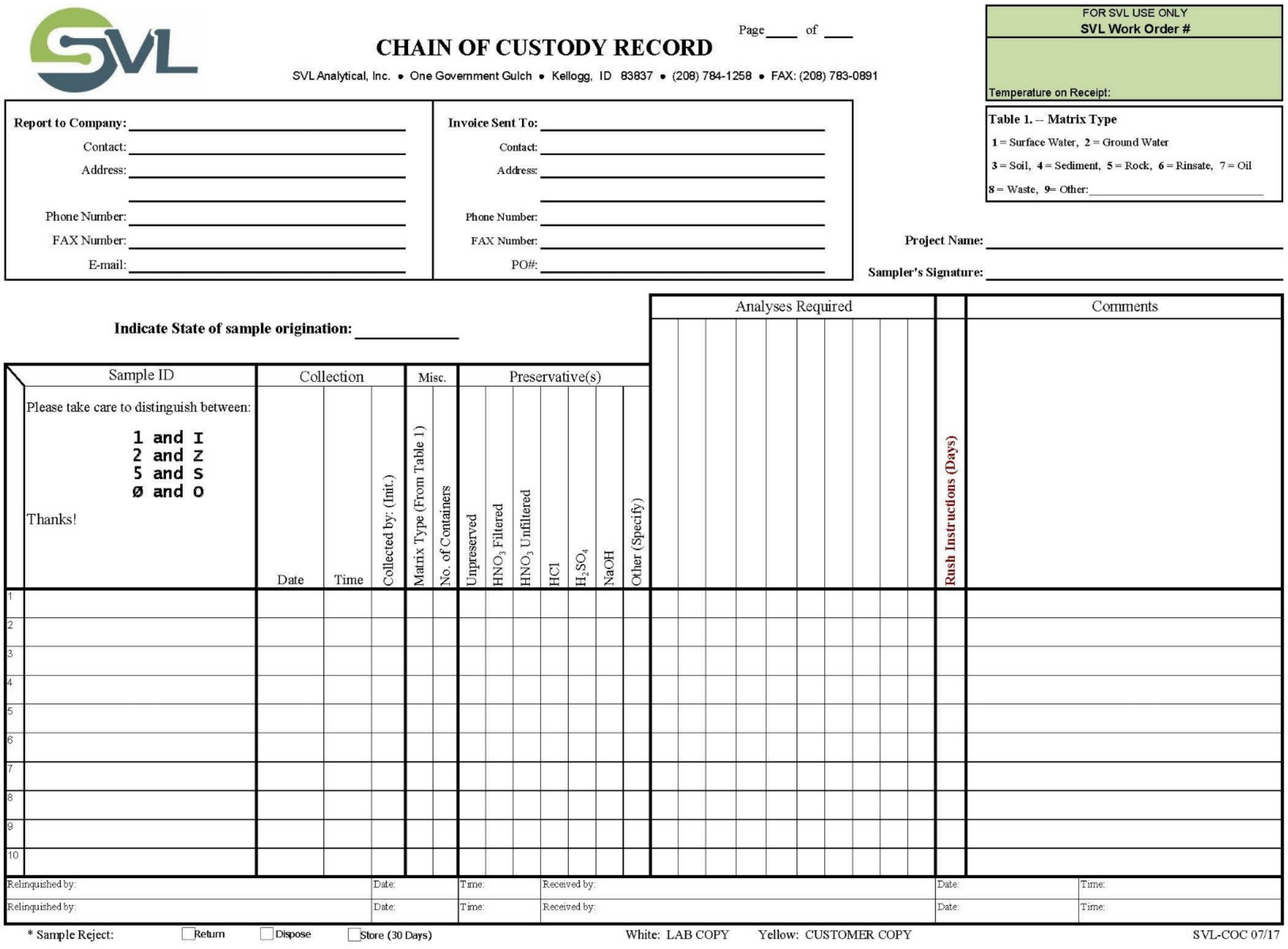
11.4 Water Quality Meters

CC&V utilizes primarily 3 types of water quality meters to collect field parameters. These meters are listed below along with the associated appendix containing calibration information:

- Aqua Troll 600 (preferred for all samples; Appendix K)
- YSI Pro (Appendix G)
- Myron Ultrameter (Appendix K)

APPENDIX A

CHAIN OF CUSTODY RECORD



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CHAIN OF CUSTODY RECORD

SVL Analytical, Inc. • One Government Gulch • Kellogg, ID 83837 • (208) 784-1258 • FAX: (208) 783-0891

FOR SVL USE ONLY
SVL Work Order #

Temperature on Receipt:

Table 1. -- Matrix Type
1 = Surface Water, 2 = Ground Water
3 = Soil, 4 = Sediment, 5 = Rock, 6 = Rinsate, 7 = Oil
8 = Waste, 9= Other: ____________

Report to Company: ____________
Contact: ____________
Address: ____________
____
Phone Number: ____________
FAX Number: ____________
E-mail: ____________

Invoice Sent To: ____________
Contact: ____________
Address: ____________
____
Phone Number: ____________
FAX Number: ____________
PO#: ____________

Project Name: ____________
Sampler's Signature: ____________

Indicate State of sample origination: ____________

	Sample ID	Collection			Misc.		Preservative(s)							Analyses Required											Comments
	Please take care to distinguish between: 1 and I 2 and Z 5 and S 0 and O Thanks!	Date	Time	Collected by: (Init.)	Matrix Type (From Table 1)	No. of Containers	Unpreserved	HNO_3 Filtered	HNO_3 Unfiltered	HCl	H_2SO_4	NaOH	Other (Specify)											Rush Instructions (Days)	
1																									
2																									
3																									
4																									
5																									
6																									
7																									
8																									
9																									
10																									

Relinquished by: Date: Time: Received by: Date: Time:
Relinquished by: Date: Time: Received by: Date: Time:

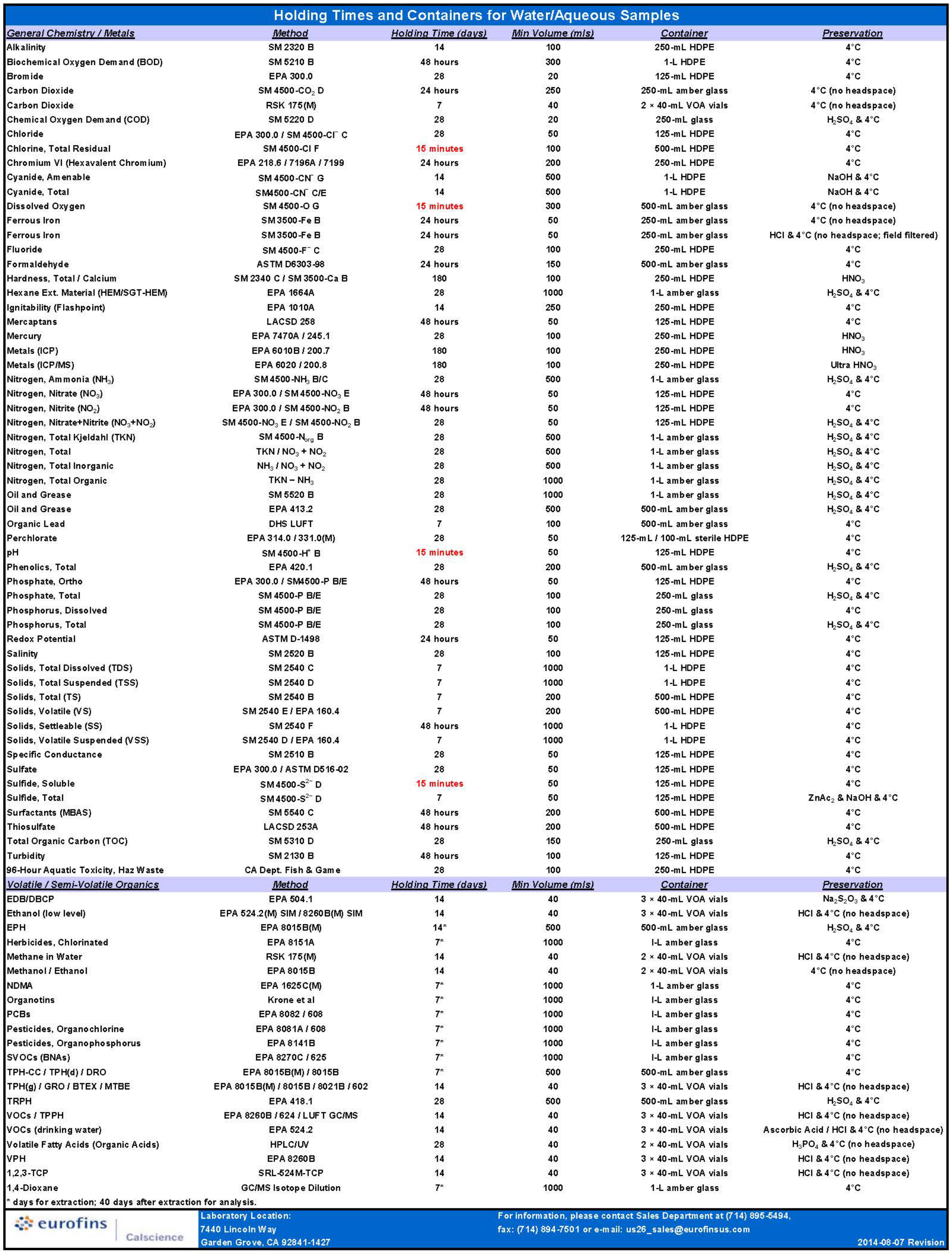
* Sample Reject: ☐ Return ☐ Dispose ☐ Store (30 Days)

White: LAB COPY Yellow: CUSTOMER COPY

SVL-COC 07/17

APPENDIX B

Field Sampling Bottle Requirements and Holding Times



Holding Times and Containers for Water/Aqueous Samples

General Chemistry / Metals	*Method*	*Holding Time (days)*	*Min Volume (mls)*	*Container*	*Preservation*
Alkalinity	SM 2320 B	14	100	250-mL HDPE	4°C
Biochemical Oxygen Demand (BOD)	SM 5210 B	48 hours	300	1-L HDPE	4°C
Bromide	EPA 300.0	28	20	125-mL HDPE	4°C
Carbon Dioxide	SM 4500-CO_2 D	24 hours	250	250-mL amber glass	4°C (no headspace)
Carbon Dioxide	RSK 175(M)	7	40	2 × 40-mL VOA vials	4°C (no headspace)
Chemical Oxygen Demand (COD)	SM 5220 D	28	20	250-mL glass	H_2SO_4 & 4°C
Chloride	EPA 300.0 / SM 4500-Cl^- C	28	50	125-mL HDPE	4°C
Chlorine, Total Residual	SM 4500-Cl F	15 minutes	100	500-mL HDPE	4°C
Chromium VI (Hexavalent Chromium)	EPA 218.6 / 7196A / 7199	24 hours	200	250-mL HDPE	4°C
Cyanide, Amenable	SM 4500-CN^- G	14	500	1-L HDPE	NaOH & 4°C
Cyanide, Total	SM4500-CN^- C/E	14	500	1-L HDPE	NaOH & 4°C
Dissolved Oxygen	SM 4500-O G	15 minutes	300	500-mL amber glass	4°C (no headspace)
Ferrous Iron	SM 3500-Fe B	24 hours	50	250-mL amber glass	4°C (no headspace)
Ferrous Iron	SM 3500-Fe B	24 hours	50	250-mL amber glass	HCl & 4°C (no headspace; field filtered)
Fluoride	SM 4500-F^- C	28	100	250-mL HDPE	4°C
Formaldehyde	ASTM D6303-98	24 hours	150	500-mL amber glass	4°C
Hardness, Total / Calcium	SM 2340 C / SM 3500-Ca B	180	100	250-mL HDPE	HNO_3
Hexane Ext. Material (HEM/SGT-HEM)	EPA 1664A	28	1000	1-L amber glass	H_2SO_4 & 4°C
Ignitability (Flashpoint)	EPA 1010A	14	250	250-mL HDPE	4°C
Mercaptans	LACSD 258	48 hours	50	125-mL HDPE	4°C
Mercury	EPA 7470A / 245.1	28	100	250-mL HDPE	HNO_3
Metals (ICP)	EPA 6010B / 200.7	180	100	250-mL HDPE	HNO_3
Metals (ICP/MS)	EPA 6020 / 200.8	180	100	250-mL HDPE	Ultra HNO_3
Nitrogen, Ammonia (NH_3)	SM 4500-NH_3 B/C	28	500	1-L amber glass	H_2SO_4 & 4°C
Nitrogen, Nitrate (NO_3)	EPA 300.0 / SM 4500-NO_3 E	48 hours	50	125-mL HDPE	4°C
Nitrogen, Nitrite (NO_2)	EPA 300.0 / SM 4500-NO_2 B	48 hours	50	125-mL HDPE	4°C
Nitrogen, Nitrate+Nitrite (NO_3+NO_2)	SM 4500-NO_3 E / SM 4500-NO_2 B	28	50	125-mL HDPE	H_2SO_4 & 4°C
Nitrogen, Total Kjeldahl (TKN)	SM 4500-N_{org} B	28	500	1-L amber glass	H_2SO_4 & 4°C
Nitrogen, Total	TKN / NO_3 + NO_2	28	500	1-L amber glass	H_2SO_4 & 4°C
Nitrogen, Total Inorganic	NH_3 / NO_3 + NO_2	28	500	1-L amber glass	H_2SO_4 & 4°C
Nitrogen, Total Organic	TKN – NH_3	28	1000	1-L amber glass	H_2SO_4 & 4°C
Oil and Grease	SM 5520 B	28	1000	1-L amber glass	H_2SO_4 & 4°C
Oil and Grease	EPA 413.2	28	500	500-mL amber glass	H_2SO_4 & 4°C
Organic Lead	DHS LUFT	7	100	500-mL amber glass	4°C
Perchlorate	EPA 314.0 / 331.0(M)	28	50	125-mL / 100-mL sterile HDPE	4°C
pH	SM 4500-H^+ B	15 minutes	50	125-mL HDPE	4°C
Phenolics, Total	EPA 420.1	28	200	500-mL amber glass	H_2SO_4 & 4°C
Phosphate, Ortho	EPA 300.0 / SM4500-P B/E	48 hours	50	125-mL HDPE	4°C
Phosphate, Total	SM 4500-P B/E	28	100	250-mL glass	H_2SO_4 & 4°C
Phosphorus, Dissolved	SM 4500-P B/E	28	100	250-mL glass	4°C
Phosphorus, Total	SM 4500-P B/E	28	100	250-mL glass	H_2SO_4 & 4°C
Redox Potential	ASTM D-1498	24 hours	50	125-mL HDPE	4°C
Salinity	SM 2520 B	28	100	125-mL HDPE	4°C
Solids, Total Dissolved (TDS)	SM 2540 C	7	1000	1-L HDPE	4°C
Solids, Total Suspended (TSS)	SM 2540 D	7	1000	1-L HDPE	4°C
Solids, Total (TS)	SM 2540 B	7	200	500-mL HDPE	4°C
Solids, Volatile (VS)	SM 2540 E / EPA 160.4	7	200	500-mL HDPE	4°C
Solids, Settleable (SS)	SM 2540 F	48 hours	1000	1-L HDPE	4°C
Solids, Volatile Suspended (VSS)	SM 2540 D / EPA 160.4	7	1000	1-L HDPE	4°C
Specific Conductance	SM 2510 B	28	50	125-mL HDPE	4°C
Sulfate	EPA 300.0 / ASTM D516-02	28	50	125-mL HDPE	4°C
Sulfide, Soluble	SM 4500-S^{2-} D	15 minutes	50	125-mL HDPE	4°C
Sulfide, Total	SM 4500-S^{2-} D	7	50	125-mL HDPE	$ZnAc_2$ & NaOH & 4°C
Surfactants (MBAS)	SM 5540 C	48 hours	200	500-mL HDPE	4°C
Thiosulfate	LACSD 253A	48 hours	200	500-mL HDPE	4°C
Total Organic Carbon (TOC)	SM 5310 D	28	150	250-mL glass	H_2SO_4 & 4°C
Turbidity	SM 2130 B	48 hours	100	125-mL HDPE	4°C
96-Hour Aquatic Toxicity, Haz Waste	CA Dept. Fish & Game	28	100	250-mL HDPE	4°C
Volatile / Semi-Volatile Organics	*Method*	*Holding Time (days)*	*Min Volume (mls)*	*Container*	*Preservation*
EDB/DBCP	EPA 504.1	14	40	3 × 40-mL VOA vials	$Na_2S_2O_3$ & 4°C
Ethanol (low level)	EPA 524.2(M) SIM / 8260B(M) SIM	14	40	3 × 40-mL VOA vials	HCl & 4°C (no headspace)
EPH	EPA 8015B(M)	14*	500	500-mL amber glass	H_2SO_4 & 4°C
Herbicides, Chlorinated	EPA 8151A	7*	1000	I-L amber glass	4°C
Methane in Water	RSK 175(M)	14	40	2 × 40-mL VOA vials	HCl & 4°C (no headspace)
Methanol / Ethanol	EPA 8015B	14	40	2 × 40-mL VOA vials	4°C (no headspace)
NDMA	EPA 1625C(M)	7*	1000	1-L amber glass	4°C
Organotins	Krone et al	7*	1000	I-L amber glass	4°C
PCBs	EPA 8082 / 608	7*	1000	I-L amber glass	4°C
Pesticides, Organochlorine	EPA 8081A / 608	7*	1000	I-L amber glass	4°C
Pesticides, Organophosphorus	EPA 8141B	7*	1000	I-L amber glass	4°C
SVOCs (BNAs)	EPA 8270C / 625	7*	1000	I-L amber glass	4°C
TPH-CC / TPH(d) / DRO	EPA 8015B(M) / 8015B	7*	500	500-mL amber glass	4°C
TPH(g) / GRO / BTEX / MTBE	EPA 8015B(M) / 8015B / 8021B / 602	14	40	3 × 40-mL VOA vials	HCl & 4°C (no headspace)
TRPH	EPA 418.1	28	500	500-mL amber glass	H_2SO_4 & 4°C
VOCs / TPPH	EPA 8260B / 624 / LUFT GC/MS	14	40	3 × 40-mL VOA vials	HCl & 4°C (no headspace)
VOCs (drinking water)	EPA 524.2	14	40	3 × 40-mL VOA vials	Ascorbic Acid / HCl & 4°C (no headspace)
Volatile Fatty Acids (Organic Acids)	HPLC/UV	28	40	2 × 40-mL VOA vials	H_3PO_4 & 4°C (no headspace)
VPH	EPA 8260B	14	40	3 × 40-mL VOA vials	HCl & 4°C (no headspace)
1,2,3-TCP	SRL-524M-TCP	14	40	3 × 40-mL VOA vials	HCl & 4°C (no headspace)
1,4-Dioxane	GC/MS Isotope Dilution	7*	1000	1-L amber glass	4°C

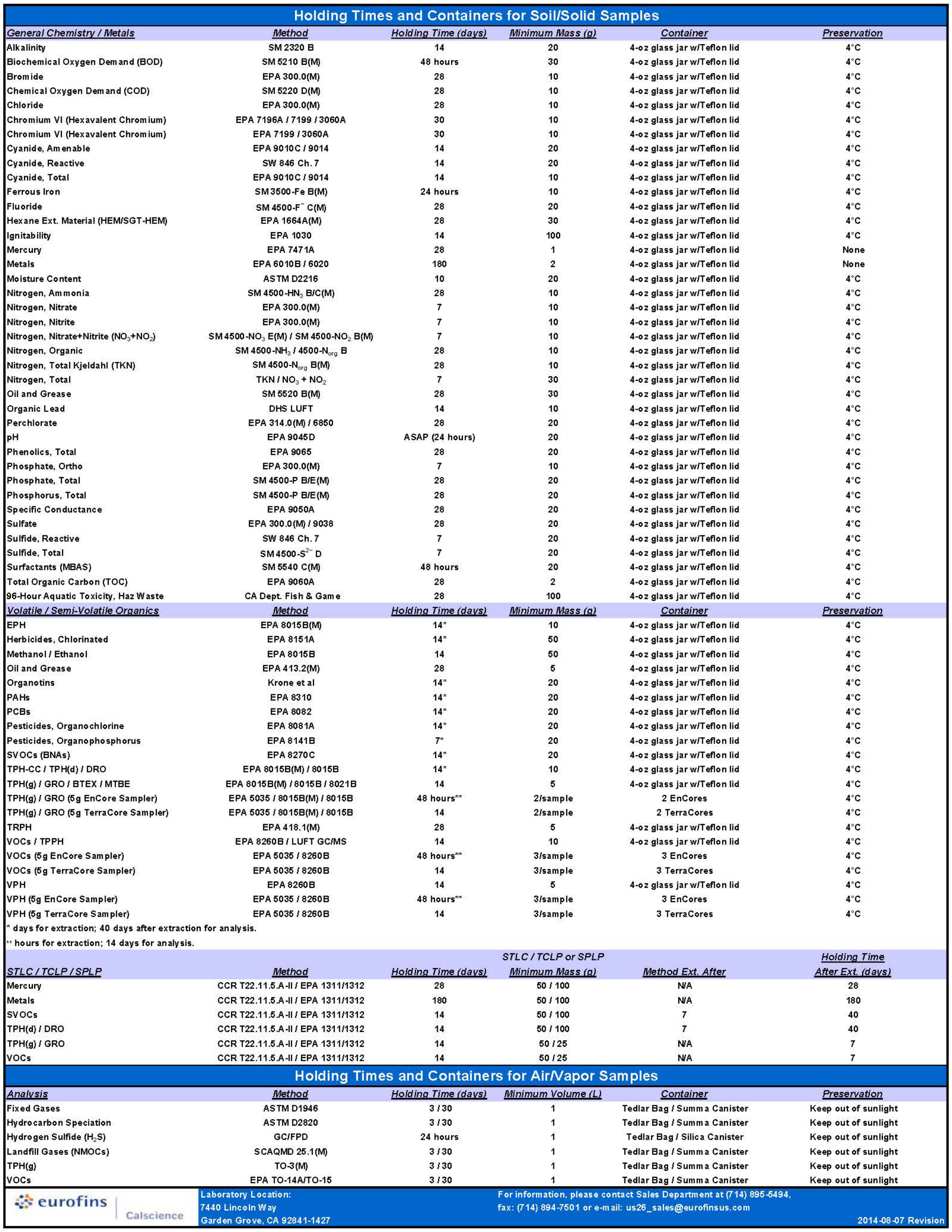
* days for extraction; 40 days after extraction for analysis.

eurofins | Calscience

Laboratory Location:
7440 Lincoln Way
Garden Grove, CA 92841-1427

For information, please contact Sales Department at (714) 895-5494,
fax: (714) 894-7501 or e-mail: us26_sales@eurofinsus.com

2014-08-07 Revision



Holding Times and Containers for Soil/Solid Samples

General Chemistry / Metals	*Method*	*Holding Time (days)*	*Minimum Mass (g)*	*Container*	*Preservation*
Alkalinity	SM 2320 B	14	20	4-oz glass jar w/Teflon lid	4°C
Biochemical Oxygen Demand (BOD)	SM 5210 B(M)	48 hours	30	4-oz glass jar w/Teflon lid	4°C
Bromide	EPA 300.0(M)	28	10	4-oz glass jar w/Teflon lid	4°C
Chemical Oxygen Demand (COD)	SM 5220 D(M)	28	10	4-oz glass jar w/Teflon lid	4°C
Chloride	EPA 300.0(M)	28	10	4-oz glass jar w/Teflon lid	4°C
Chromium VI (Hexavalent Chromium)	EPA 7196A / 7199 / 3060A	30	10	4-oz glass jar w/Teflon lid	4°C
Chromium VI (Hexavalent Chromium)	EPA 7199 / 3060A	30	10	4-oz glass jar w/Teflon lid	4°C
Cyanide, Amenable	EPA 9010C / 9014	14	20	4-oz glass jar w/Teflon lid	4°C
Cyanide, Reactive	SW 846 Ch. 7	14	20	4-oz glass jar w/Teflon lid	4°C
Cyanide, Total	EPA 9010C / 9014	14	10	4-oz glass jar w/Teflon lid	4°C
Ferrous Iron	SM 3500-Fe B(M)	24 hours	10	4-oz glass jar w/Teflon lid	4°C
Fluoride	SM 4500-F^- C(M)	28	20	4-oz glass jar w/Teflon lid	4°C
Hexane Ext. Material (HEM/SGT-HEM)	EPA 1664A(M)	28	30	4-oz glass jar w/Teflon lid	4°C
Ignitability	EPA 1030	14	100	4-oz glass jar w/Teflon lid	4°C
Mercury	EPA 7471A	28	1	4-oz glass jar w/Teflon lid	None
Metals	EPA 6010B / 6020	180	2	4-oz glass jar w/Teflon lid	None
Moisture Content	ASTM D2216	10	20	4-oz glass jar w/Teflon lid	4°C
Nitrogen, Ammonia	SM 4500-HN_3 B/C(M)	28	10	4-oz glass jar w/Teflon lid	4°C
Nitrogen, Nitrate	EPA 300.0(M)	7	10	4-oz glass jar w/Teflon lid	4°C
Nitrogen, Nitrite	EPA 300.0(M)	7	10	4-oz glass jar w/Teflon lid	4°C
Nitrogen, Nitrate+Nitrite (NO_3+NO_2)	SM 4500-NO_3 E(M) / SM 4500-NO_2 B(M)	7	10	4-oz glass jar w/Teflon lid	4°C
Nitrogen, Organic	SM 4500-NH_3 / 4500-N_{org} B	28	10	4-oz glass jar w/Teflon lid	4°C
Nitrogen, Total Kjeldahl (TKN)	SM 4500-N_{org} B(M)	28	10	4-oz glass jar w/Teflon lid	4°C
Nitrogen, Total	TKN / NO_3 + NO_2	7	30	4-oz glass jar w/Teflon lid	4°C
Oil and Grease	SM 5520 B(M)	28	30	4-oz glass jar w/Teflon lid	4°C
Organic Lead	DHS LUFT	14	10	4-oz glass jar w/Teflon lid	4°C
Perchlorate	EPA 314.0(M) / 6850	28	20	4-oz glass jar w/Teflon lid	4°C
pH	EPA 9045D	ASAP (24 hours)	20	4-oz glass jar w/Teflon lid	4°C
Phenolics, Total	EPA 9065	28	20	4-oz glass jar w/Teflon lid	4°C
Phosphate, Ortho	EPA 300.0(M)	7	10	4-oz glass jar w/Teflon lid	4°C
Phosphate, Total	SM 4500-P B/E(M)	28	20	4-oz glass jar w/Teflon lid	4°C
Phosphorus, Total	SM 4500-P B/E(M)	28	20	4-oz glass jar w/Teflon lid	4°C
Specific Conductance	EPA 9050A	28	20	4-oz glass jar w/Teflon lid	4°C
Sulfate	EPA 300.0(M) / 9038	28	20	4-oz glass jar w/Teflon lid	4°C
Sulfide, Reactive	SW 846 Ch. 7	7	20	4-oz glass jar w/Teflon lid	4°C
Sulfide, Total	SM 4500-S^{2-} D	7	20	4-oz glass jar w/Teflon lid	4°C
Surfactants (MBAS)	SM 5540 C(M)	48 hours	20	4-oz glass jar w/Teflon lid	4°C
Total Organic Carbon (TOC)	EPA 9060A	28	2	4-oz glass jar w/Teflon lid	4°C
96-Hour Aquatic Toxicity, Haz Waste	CA Dept. Fish & Game	28	100	4-oz glass jar w/Teflon lid	4°C
Volatile / Semi-Volatile Organics	*Method*	*Holding Time (days)*	*Minimum Mass (g)*	*Container*	*Preservation*
EPH	EPA 8015B(M)	14*	10	4-oz glass jar w/Teflon lid	4°C
Herbicides, Chlorinated	EPA 8151A	14*	50	4-oz glass jar w/Teflon lid	4°C
Methanol / Ethanol	EPA 8015B	14	50	4-oz glass jar w/Teflon lid	4°C
Oil and Grease	EPA 413.2(M)	28	5	4-oz glass jar w/Teflon lid	4°C
Organotins	Krone et al	14*	20	4-oz glass jar w/Teflon lid	4°C
PAHs	EPA 8310	14*	20	4-oz glass jar w/Teflon lid	4°C
PCBs	EPA 8082	14*	20	4-oz glass jar w/Teflon lid	4°C
Pesticides, Organochlorine	EPA 8081A	14*	20	4-oz glass jar w/Teflon lid	4°C
Pesticides, Organophosphorus	EPA 8141B	7*	20	4-oz glass jar w/Teflon lid	4°C
SVOCs (BNAs)	EPA 8270C	14*	20	4-oz glass jar w/Teflon lid	4°C
TPH-CC / TPH(d) / DRO	EPA 8015B(M) / 8015B	14*	10	4-oz glass jar w/Teflon lid	4°C
TPH(g) / GRO / BTEX / MTBE	EPA 8015B(M) / 8015B / 8021B	14	5	4-oz glass jar w/Teflon lid	4°C
TPH(g) / GRO (5g EnCore Sampler)	EPA 5035 / 8015B(M) / 8015B	48 hours**	2/sample	2 EnCores	4°C
TPH(g) / GRO (5g TerraCore Sampler)	EPA 5035 / 8015B(M) / 8015B	14	2/sample	2 TerraCores	4°C
TRPH	EPA 418.1(M)	28	5	4-oz glass jar w/Teflon lid	4°C
VOCs / TPPH	EPA 8260B / LUFT GC/MS	14	10	4-oz glass jar w/Teflon lid	4°C
VOCs (5g EnCore Sampler)	EPA 5035 / 8260B	48 hours**	3/sample	3 EnCores	4°C
VOCs (5g TerraCore Sampler)	EPA 5035 / 8260B	14	3/sample	3 TerraCores	4°C
VPH	EPA 8260B	14	5	4-oz glass jar w/Teflon lid	4°C
VPH (5g EnCore Sampler)	EPA 5035 / 8260B	48 hours**	3/sample	3 EnCores	4°C
VPH (5g TerraCore Sampler)	EPA 5035 / 8260B	14	3/sample	3 TerraCores	4°C

* days for extraction; 40 days after extraction for analysis.

** hours for extraction; 14 days for analysis.

STLC / TCLP / SPLP	*Method*	*Holding Time (days)*	*STLC / TCLP or SPLP Minimum Mass (g)*	*Method Ext. After*	*Holding Time After Ext. (days)*
Mercury	CCR T22.11.5.A-II / EPA 1311/1312	28	50 / 100	N/A	28
Metals	CCR T22.11.5.A-II / EPA 1311/1312	180	50 / 100	N/A	180
SVOCs	CCR T22.11.5.A-II / EPA 1311/1312	14	50 / 100	7	40
TPH(d) / DRO	CCR T22.11.5.A-II / EPA 1311/1312	14	50 / 100	7	40
TPH(g) / GRO	CCR T22.11.5.A-II / EPA 1311/1312	14	50 / 25	N/A	7
VOCs	CCR T22.11.5.A-II / EPA 1311/1312	14	50 / 25	N/A	7

Holding Times and Containers for Air/Vapor Samples

Analysis	*Method*	*Holding Time (days)*	*Minimum Volume (L)*	*Container*	*Preservation*
Fixed Gases	ASTM D1946	3 / 30	1	Tedlar Bag / Summa Canister	Keep out of sunlight
Hydrocarbon Speciation	ASTM D2820	3 / 30	1	Tedlar Bag / Summa Canister	Keep out of sunlight
Hydrogen Sulfide (H_2S)	GC/FPD	24 hours	1	Tedlar Bag / Silica Canister	Keep out of sunlight
Landfill Gases (NMOCs)	SCAQMD 25.1(M)	3 / 30	1	Tedlar Bag / Summa Canister	Keep out of sunlight
TPH(g)	TO-3(M)	3 / 30	1	Tedlar Bag / Summa Canister	Keep out of sunlight
VOCs	EPA TO-14A/TO-15	3 / 30	1	Tedlar Bag / Summa Canister	Keep out of sunlight

eurofins | Calscience

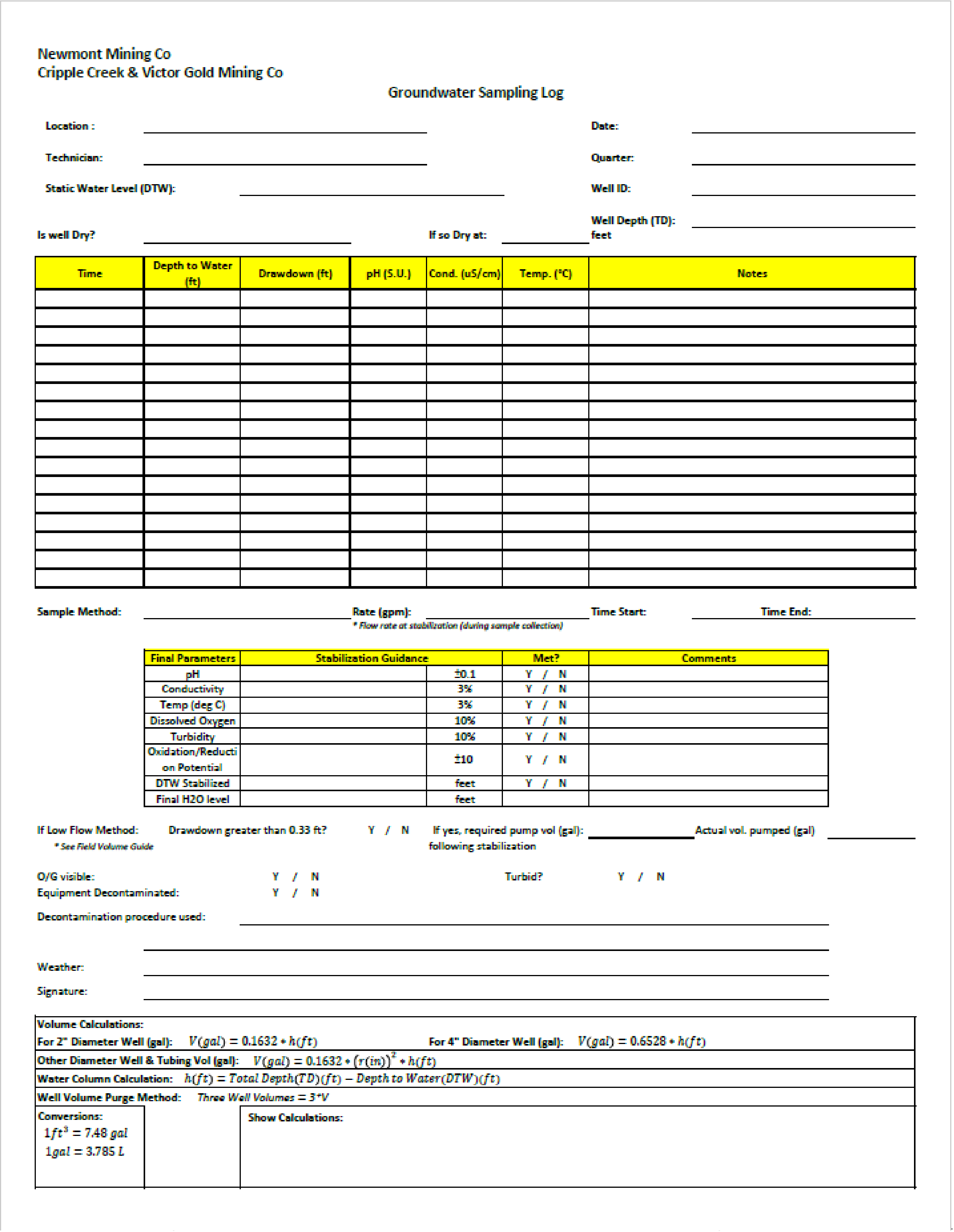
Laboratory Location:
7440 Lincoln Way
Garden Grove, CA 92841-1427

For information, please contact Sales Department at (714) 895-5494,
fax: (714) 894-7501 or e-mail: us26_sales@eurofinsus.com

2014-08-07 Revision

APPENDIX C

Example Field Sample Record Sheets



Newmont Mining Co
Cripple Creek & Victor Gold Mining Co

Groundwater Sampling Log

Location : ______________________ **Date:** ______________________

Technician: ______________________ **Quarter:** ______________________

Static Water Level (DTW): ______________________ **Well ID:** ______________________

Well Depth (TD): ______________________ feet

Is well Dry? ______________________ **If so Dry at:** ______________________

Time	Depth to Water (ft)	Drawdown (ft)	pH (S.U.)	Cond. (uS/cm)	Temp. (°C)	Notes

Sample Method: ______________________ **Rate (gpm):** ______________________ **Time Start:** ______________________ **Time End:** ______________________

** Flow rate at stabilization (during sample collection)*

Final Parameters	Stabilization Guidance		Met?	Comments
pH		±0.1	Y / N	
Conductivity		3%	Y / N	
Temp (deg C)		3%	Y / N	
Dissolved Oxygen		10%	Y / N	
Turbidity		10%	Y / N	
Oxidation/Reduction Potential		±10	Y / N	
DTW Stabilized		feet	Y / N	
Final H2O level		feet		

If Low Flow Method: Drawdown greater than 0.33 ft? Y / N If yes, required pump vol (gal): ______________________ Actual vol. pumped (gal) ______________________

** See Field Volume Guide* following stabilization

O/G visible: Y / N Turbid? Y / N

Equipment Decontaminated: Y / N

Decontamination procedure used: ______________________

____

Weather: ______________________

Signature: ______________________

Volume Calculations:

For 2" Diameter Well (gal): $V(gal) = 0.1632 * h(ft)$ **For 4" Diameter Well (gal):** $V(gal) = 0.6528 * h(ft)$

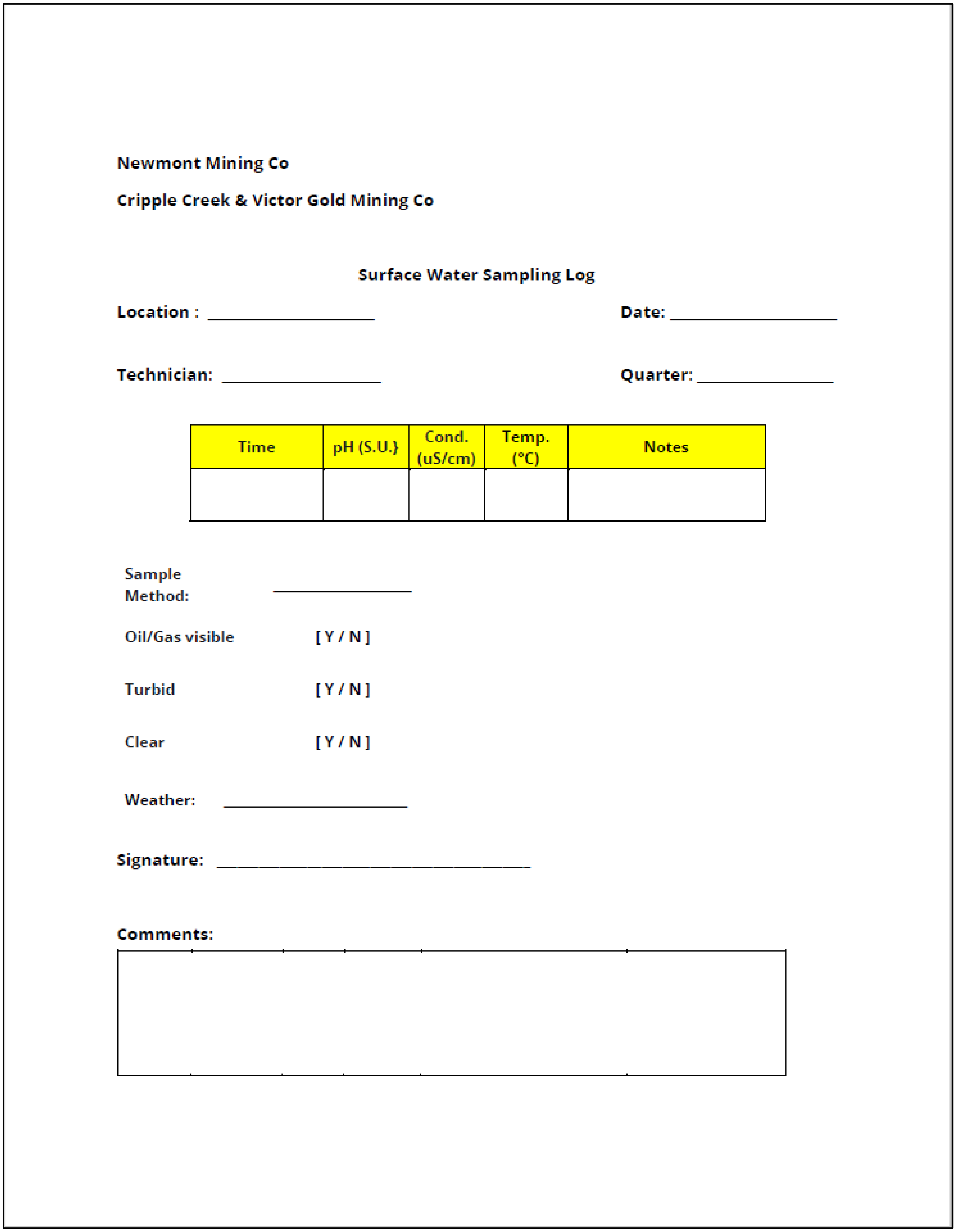
Other Diameter Well & Tubing Vol (gal): $V(gal) = 0.1632 * (r(in))^2 * h(ft)$

Water Column Calculation: $h(ft) = Total\ Depth(TD)(ft) - Depth\ to\ Water(DTW)(ft)$

Well Volume Purge Method: *Three Well Volumes = 3*V*

Conversions:
$1 ft^3 = 7.48\ gal$
$1 gal = 3.785\ L$

Show Calculations:



Newmont Mining Co

Cripple Creek & Victor Gold Mining Co

Surface Water Sampling Log

Location : ____________________ **Date:** ____________________

Technician: ___________________ **Quarter:** ________________

Time	pH (S.U.)	Cond. (uS/cm)	Temp. (°C)	Notes

Sample Method: ________________

Oil/Gas visible [Y / N]

Turbid [Y / N]

Clear [Y / N]

Weather: ______________________

Signature: ______________________________________

Comments:

Low-Flow Test Report:

Test Date / Time: 3/20/2024 2:36:24 PM
Project:
Operator Name:

Location Name: Device Location **Initial Depth to Water: 93.15 ft**	**Estimated Total Volume Pumped: 12.784 gal** **Flow Cell Volume: 130 ml** **Final Flow Rate: 0.34 gal/min** **Final Draw Down: 2.22 ft**	**Instrument Used: Aqua TROLL 600** **Serial Number: 1109809**

Test Notes:
Grab sample, suspected broken casing

Low-Flow Readings:

Date Time	Elapsed Time	pH	Temperature	Specific Conductivity	RDO Concentration	Turbidity	ORP	Depth to Water	Flow
		+/- 0.1	+/- 3 %	+/- 3 %	+/- 10 %	+/- 10 %	+/- 10 %	+/- 0.33	
3/20/2024 2:36 PM	00:00	6.75 pH	7.85 °C	403.47 µS/cm	1.69 mg/L	43.88 NTU	9.1 mV	94.90 ft	0.34 gal/min
3/20/2024 2:37 PM	01:22	6.75 pH	6.56 °C	416.97 µS/cm	0.40 mg/L	43.68 NTU	50.5 mV	94.92 ft	0.34 gal/min
3/20/2024 2:42 PM	06:22	6.61 pH	6.08 °C	418.16 µS/cm	0.25 mg/L	50.58 NTU	104.2 mV	95.04 ft	0.34 gal/min
3/20/2024 2:47 PM	11:22	6.54 pH	6.25 °C	413.27 µS/cm	0.22 mg/L	59.01 NTU	95.7 mV	95.15 ft	0.34 gal/min
3/20/2024 2:52 PM	16:22	6.52 pH	6.40 °C	407.51 µS/cm	0.18 mg/L	61.13 NTU	77.7 mV	95.18 ft	0.34 gal/min
3/20/2024 2:54 PM	17:36	6.51 pH	6.37 °C	409.28 µS/cm	0.19 mg/L	59.00 NTU	83.9 mV	95.18 ft	0.34 gal/min
3/20/2024 2:59 PM	22:36	6.51 pH	6.30 °C	406.32 µS/cm	0.16 mg/L	59.11 NTU	112.1 mV	95.28 ft	0.34 gal/min
3/20/2024 3:04 PM	27:36	6.51 pH	6.39 °C	401.73 µS/cm	0.16 mg/L	60.35 NTU	83.7 mV	95.35 ft	0.34 gal/min
3/20/2024 3:09 PM	32:36	6.52 pH	6.42 °C	395.79 µS/cm	0.14 mg/L	60.92 NTU	57.2 mV	95.37 ft	0.34 gal/min
3/20/2024 3:14 PM	37:36	6.59 pH	7.44 °C	389.70 µS/cm	0.15 mg/L	67.32 NTU	79.1 mV	95.37 ft	0.34 gal/min

Samples

Sample ID:	Description:
GVMW15A	

Created using VuSitu from In-Situ, Inc.

Appendix D
Reagent Water Specifications

REAGENT WATER SPECIFICATIONS

Quality Parameter	Type I	Type II	Type III
Bacteria, CFU/Ml	10	1000	NA
pH	NA	NA	8-May
Resistivity, megohm-cm at 25□ C	> 10	> 1	0.11
Resistivity, megohm-cm at 25□ C	> 10	> 1	0.1
Conductivity, umho/cm at 25□ C	< 0.1	1	10
SiO_2, mg/L	< 0.05	< 0.1	< 1
Total Solids, mg/L	0.1	1	5
Total oxidizable organic carbon, mg/l	< 0.05	< 0.2	< 1

* NA = not applicable

Reference - Standard Methods for the Examination of Water and Wastewater, 17th Edition, 1989.

APPENDIX E

WELL EVACUATION CALCULATION

WELL EVACUATION CALCULATION - EXAMPLE

1. Measure depth to water from top of casing (TOC) prior to purging.
2. Begin purging well.
3. Determine well casing volume.

Total depth of well (TD) -depth to water (DTW) = total height of water in casing (H).

Example: TD = 66.60'
DTW = 46.15'
H = 20.45'

Volume in cubic feet (Vc) = 3.14 x (radius of well)^2 x H

Example: with a 4" casing, radius = 2" = 0.167'

Vc = 3.14 x (0.167')^2 x 20.45'
Vc = 0.09 square feet x 20.45 feet
Vc = 1.8 cubic feet

4. Convert cubic feet to gallons.

Cubic feet x 7.48 = gallons
Example: Vg = 1.8 cubic feet x 7.48 = 13.5 gallons

5. Three well volumes must be evacuated:

Example: Vw_3 = 3 x 13.5 gallons = 40.4 gallons

6. Hence, to evacuate three well volumes in the above example, 40.4 gallons need to be purged before sampling.

Since all values in the above calculations are constant except for the height of water in casing (H), the constant values may be pre-calculated to simplify well volume determinations.
Thus:
For a 2" well, three well volumes (Vw_3) = H x 0.5
For a 4" well, three well volumes (Vw_3) = H x 2

APPENDIX F

Low-Flow Purge Volume Calculation

Groundwater Well - Field Volume Guide

Volume Equation:	$V(gal) = 0.1632 \cdot (r(in))^2 \cdot \Delta(ft)$
Delta (Δ) (ft):	$\Delta(ft) = Stabilized\ Depth\ to\ Water(ft) - Initial\ Depth\ to\ Water(ft)$

Required Pumping Volume:	$Pumping\ Volume\ (gal) = \Delta V(gal) + Tubing\ Volume\ (gal)$

Diameter (in)	Volume (gal)
1"	$V(gal) = 0.0408 \cdot \Delta(ft)$
2"	$V(gal) = 0.1632 \cdot \Delta(ft)$
3"	$V(gal) = 0.3672 \cdot \Delta(ft)$
4"	$V(gal) = 0.6528 \cdot \Delta(ft)$
5"	$V(gal) = 1.02 \cdot \Delta(ft)$

ΔV - Volume b/w initial level and stabilized level (gal)

Delta (Δ) (ft)	1" Well	2" Well	3" Well	4" Well	5" Well
0.1	0.0	0.0	0.0	0.1	0.1
0.3	0.0	0.0	0.1	0.2	0.3
0.5	0.0	0.1	0.2	0.3	0.5
0.7	0.0	0.1	0.3	0.5	0.7
0.9	0.0	0.1	0.3	0.6	0.9
1.1	0.0	0.2	0.4	0.7	1.1
1.3	0.1	0.2	0.5	0.8	1.3
1.5	0.1	0.2	0.6	1.0	1.5
1.7	0.1	0.3	0.6	1.1	1.7
1.9	0.1	0.3	0.7	1.2	1.9
2.1	0.1	0.3	0.8	1.4	2.1
2.3	0.1	0.4	0.8	1.5	2.3
2.5	0.1	0.4	0.9	1.6	2.6
2.7	0.1	0.4	1.0	1.8	2.8
2.9	0.1	0.5	1.1	1.9	3.0
3.1	0.1	0.5	1.1	2.0	3.2
3.3	0.1	0.5	1.2	2.2	3.4
3.5	0.1	0.6	1.3	2.3	3.6
3.7	0.2	0.6	1.4	2.4	3.8
3.9	0.2	0.6	1.4	2.5	4.0
4.1	0.2	0.7	1.5	2.7	4.2
4.3	0.2	0.7	1.6	2.8	4.4
4.5	0.2	0.7	1.7	2.9	4.6
4.7	0.2	0.8	1.7	3.1	4.8
4.9	0.2	0.8	1.8	3.2	5.0
5.1	0.2	0.8	1.9	3.3	5.2
5.3	0.2	0.9	1.9	3.5	5.4
5.5	0.2	0.9	2.0	3.6	5.6
5.7	0.2	0.9	2.1	3.7	5.8
5.9	0.2	1.0	2.2	3.9	6.0
6.1	0.2	1.0	2.2	4.0	6.2
6.3	0.3	1.0	2.3	4.1	6.4
6.5	0.3	1.1	2.4	4.2	6.6
6.7	0.3	1.1	2.5	4.4	6.8
6.9	0.3	1.1	2.5	4.5	7.0
7.1	0.3	1.2	2.6	4.6	7.2
7.3	0.3	1.2	2.7	4.8	7.4
7.5	0.3	1.2	2.8	4.9	7.7
7.7	0.3	1.3	2.8	5.0	7.9
7.9	0.3	1.3	2.9	5.2	8.1
8.1	0.3	1.3	3.0	5.3	8.3
8.3	0.3	1.4	3.0	5.4	8.5
8.5	0.3	1.4	3.1	5.5	8.7
8.7	0.4	1.4	3.2	5.7	8.9
8.9	0.4	1.5	3.3	5.8	9.1
9.1	0.4	1.5	3.3	5.9	9.3
9.3	0.4	1.5	3.4	6.1	9.5
9.5	0.4	1.6	3.5	6.2	9.7
9.7	0.4	1.6	3.6	6.3	9.9
9.9	0.4	1.6	3.6	6.5	10.1
10.1	0.4	1.6	3.7	6.6	10.3

APPENDIX G
YSI Pro Water Quality Meter Calibration

PURPOSE

CC&V uses the YSI Pro Plus water quality meter to collect water quality data that is submitted to regulatory agencies. It is the responsibility of the user to properly calibrate the probe before each use and keep an accurate record of each calibration.

SCOPE

The YSI pro plus is to be calibrated prior to use for the day and details of the calibration are to be recorded on the calibration log sheet.

YSI CALIBRATION

Dissolved Oxygen Calibration

Moisten the plastic cup by adding a small amount of clean water (1/8 inch) in the plastic storage cup or by moistening the sponge in the bottom of the cup. Make sure there are no water droplets on the DO membrane or temperature sensor. Then install the cup over the sensors screw it on the cable and then disengage one or two threads to ensure atmospheric venting. Make sure the DO and temperature sensors are not immersed in water. Turn the instrument on and wait approximately 5 to 15 minutes for the storage container to become completely saturated and to allow the sensors to stabilize.

It is not necessary to calibrate in both % and mg/L or ppm. Calibrating in % will simultaneously calibrate mg/L and ppm and vice versa.

Press 'Cal' Highlight Probe ID
Highlight DO % and press enter to confirm.
Wait for the temperature and DO% values under "Actual Readings" to stabilize.
Then highlight Accept Calibration and press enter to calibrate. Or, press Esc to cancel the calibration.

Specific Conductance Calibration

Press 'Cal' Highlight Probe ID.
After selecting the Probe ID, highlight Conductivity and press enter. Highlight the desired calibration method; Sp. Conductance. Remove sponge from cup. Place the sensor into a fresh, traceable conductivity calibration solution. The solution must cover the holes of the conductivity sensor that are closest to the cable.

Ensure the entire conductivity sensor is submerged in the solution or the instrument will read approximately of half the expected value!

Choose the units in either SPC-us/cm and press enter.

Highlight Calibration value and press enter to input the value of the calibration standard. Then, once the temperature and conductivity readings stabilize, highlight Accept Calibration and press enter. Or, press Esc to cancel the calibration

pH Calibration

Press 'Cal'. Highlight Probe ID. After selecting your or Probe ID, highlight ISE (pH) and press enter. The message line will show the instrument is "Ready for point 1". The pH calibration allows up to six calibration points. Calibration for CCV purposes will be a 3 point calibration (4, 7, and 10). Place the sensor in a pH 7 buffer solution. The instrument should automatically recognize the buffer value and display it at the top of the calibration screen. If the calibration value is incorrect, the auto buffer recognition setting in the Sensor Setup menu may be incorrect. If necessary, highlight the Calibration Value and press enter to input the correct buffer value. Once the pH and temperature readings stabilize, highlight Accept Calibration and press enter to accept the first calibration point. The message line will then display "Ready for point 2".

To continue with the 2nd point, place the sensor in the second buffer solution (pH4). The instrument should automatically recognize the second buffer value (pH4) and display it at the top of the screen. If necessary, highlight the Calibration Value and press enter to input the correct buffer value. Once the pH and temperature readings stabilize, highlight Accept Calibration and press enter to confirm the second calibration point.

The message line will then display 'Ready for point 3" and you can continue with the 3rd calibration point (pH10) as detailed in previous steps for first and second calibration points.

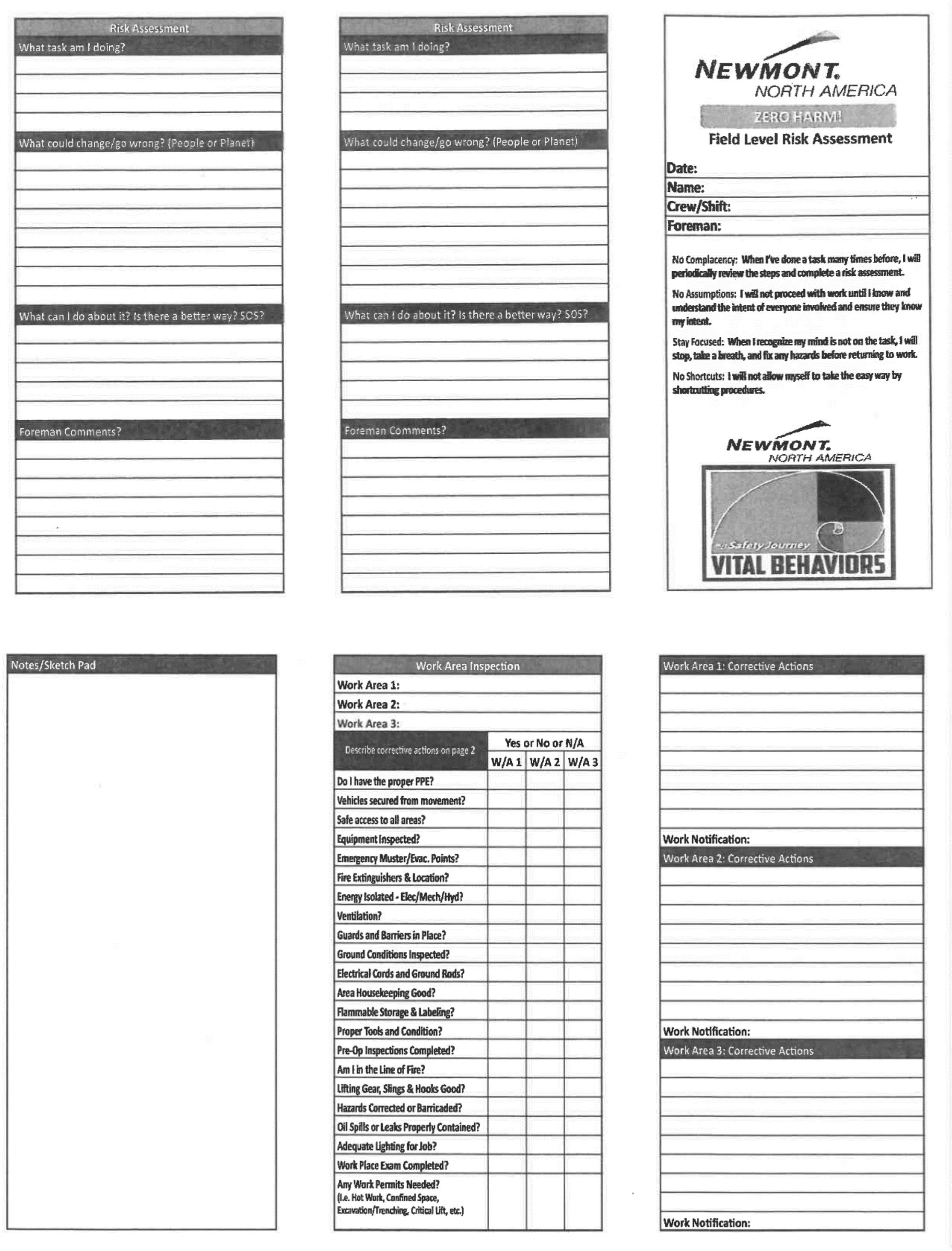
Press 'Cal' to complete the calibration.

Calibration Finalization

Upon completing the calibrations prior to field use, return probes to pH 4 storage solution. pH 4 is the recommended storage solution to prevent the dehydration of the pH probe. While using YSI, place pH 4 storage solution in the storage solution container that is maintained in case, to be used for continued storage at days end. Insure you have recorded the required values on the YSI Pro Plus calibration log.

APPENDIX H

Field Level Risk Assessment



Risk Assessment

What task am I doing?

What could change/go wrong? (People or Planet)

What can I do about it? Is there a better way? SOS?

Foreman Comments?

Risk Assessment

What task am I doing?

What could change/go wrong? (People or Planet)

What can I do about it? Is there a better way? SOS?

Foreman Comments?

NEWMONT.
NORTH AMERICA
ZERO HARM!
Field Level Risk Assessment

Date:
Name:
Crew/Shift:
Foreman:

No Complacency: When I've done a task many times before, I will periodically review the steps and complete a risk assessment.

No Assumptions: I will not proceed with work until I know and understand the intent of everyone involved and ensure they know my intent.

Stay Focused: When I recognize my mind is not on the task, I will stop, take a breath, and fix any hazards before returning to work.

No Shortcuts: I will not allow myself to take the easy way by shortcutting procedures.

NEWMONT.
NORTH AMERICA
...a Safety Journey
VITAL BEHAVIORS

Notes/Sketch Pad

Work Area Inspection

Work Area 1:
Work Area 2:
Work Area 3:

Describe corrective actions on page 2	Yes or No or N/A		
	W/A 1	W/A 2	W/A 3
Do I have the proper PPE?			
Vehicles secured from movement?			
Safe access to all areas?			
Equipment Inspected?			
Emergency Muster/Evac. Points?			
Fire Extinguishers & Location?			
Energy Isolated - Elec/Mech/Hyd?			
Ventilation?			
Guards and Barriers in Place?			
Ground Conditions Inspected?			
Electrical Cords and Ground Rods?			
Area Housekeeping Good?			
Flammable Storage & Labeling?			
Proper Tools and Condition?			
Pre-Op Inspections Completed?			
Am I in the Line of Fire?			
Lifting Gear, Slings & Hooks Good?			
Hazards Corrected or Barricaded?			
Oil Spills or Leaks Properly Contained?			
Adequate Lighting for Job?			
Work Place Exam Completed?			
Any Work Permits Needed? (I.e. Hot Work, Confined Space, Excavation/Trenching, Critical Lift, etc.)			

Work Area 1: Corrective Actions

Work Notification:

Work Area 2: Corrective Actions

Work Notification:

Work Area 3: Corrective Actions

Work Notification:

APPENDIX I

CC&V Surface Water Flume & Weir Calibration SOP

PURPOSE

Cripple Creek & Victor utilizes flumes and weirs to measure surface water flow, this instruction is to aid in the inspections, and confirmation of the accuracy of the flumes and weirs.

SCOPE

Calibrations of the flumes and weir are to be conducted on an annual basis and recorded in the proper sharedrive location.

PPE

Safety Toe boots
Safety glasses
Hard hat
High-visibility vest

EQUIPMENT

Bucket
Stopwatch
Staff
Marsh-McBirney Flo-mate
Level

Flume Calibration

To confirm flume is level, use a long enough level to span the support brackets of the flume length wise, and across the width at the inlet, and the outlet. Check level regardless of absent flow.

If flume is out of level use the adjustment all threads and adjust flume to level. There are four adjustment rods for adjusting level. Some excavating may be necessary to access the adjustments.

If flow through flume is greater than 0.3 you may use flo-mate to calibrate flume. If flow is less than 0.3 use bucket and stop watch method (See Weir Calibration)

To calibrate flow through the flume, attach flo-mate reader to staff. Note the depth of water on flume staff.

Place staff and flo-mate in flume with instrument at 30% total depth from bottom. Instrument will provide a velocity measurement.

Calculate flow:
Depth x width x velocity = flow

Compare instrument measurement with known value attained from water depth in flume. Values should be with-in 10% of each other. Document values on the flume calibration log. If flow is not sufficient to use flo-mate, use calibrated bucket and stop watch to measure flow.

Weir Calibration

Use calibrated bucket to collect water, use stop watch to time the collection of water to calculate flow.
Volume of bucket / Collection time= flow x 60= GPM
Ex. 1 gallon 5.0 seconds
1/5 x60 =12 GPM

Compare calculated flow to flow reading from weir.

APPENDIX J

CC&V Sample Shipping SOP

CC&V – Sample Prep & Shipping SOP

SAMPLE PREP & SHIPPING SUMMARY

The following inventory of prepared sample bottles must be continually kept up:

1) **Cyanide Bottles**: green labels, brown 125 ml bottles preserved with 0.2 ml of Sodium Hydroxide ~60 bottles

2) **Metals Bottles:** red labels, white 250 ml bottles preserved with 1.5 ml of Nitric Acid ~ 60 bottles

3) **Nitrogen Bottles**: yellow labels, white 250 ml bottles preserved with 0.5 ml of Sulfuric Acid ~40 bottles

4) **Mineral Bottles:** black labels, 500 ml bottles non-preserved ~ 60 bottles

5) **Hexavalent Chromium Kit:** green labels, 250 mL bottles preserved with buffer, NaOH for pH adjustment, and pH strips ~ 20 bottles

6) **Sulfide Bottles:** purple labels, 250 mL bottles preserved with Zinc Acetate NaOH ~ 20 bottles

5) **Verify expiration dates** on reagent used in bottle preservation

Before running too low on bottles or preservatives order from SVL. Always confirm sample supplies before each new quarter to ensure sufficient supply for quarterly compliance sampling.

ALWAYS WEAR NITRILE GLOVES AND SAFETY GLASSES WHEN PRESERVING BOTTLES!!!!

Shipping Samples

Check with staff daily if there will be samples ready to ship out at the end of the day, if there are samples to ship follow the shipping instructions:

1) Match COC's to bottles in the refrigerator
2) Sign and date the COC's
3) Make copies of COC's, place original in plastic bag – these will go with the cooler with samples. Place copies in red file folder for reference.
4) Load up samples, COC's, and ice into "five-day" cooler.
5) Sign and date Custody seal and place vertically across lid and cooler.
6) Tape up cooler lengthwise and width wise
7) Take cooler and mailing label up to the Post Office no later than 4:15 p.m.
8) Get mailing label sticker with the tracking number and place it on the copies of the COC's
9) Keep freezer stocked full of ice!!

Procedures for Water Quality Sample Shipments.

1) Any water quality samples collected from Monday thru Thursday should be shipped either before or on Thursday.
a) CDPS Water Quality Samples are to be shipped the day the sample was collected if time allows, otherwise they must be shipped the next day.
b) Ground & Surface Water Samples are to be shipped either on the day the sample was collected or the next day.
c) VLF Water Quality Samples are to be shipped either on the day the sample was collected or within two days.
2) Any water quality sample collected on Friday may sit in the refrigerator until the following Monday, but must be shipped on that Monday.
a) No water quality samples are to sit in refrigerator over a weekend, without prior approval.

APPENDIX K

Myron L II Ultrameter Calibration procedure

PURPOSE

The Myron L II is used to collect data that is submitted to regulatory agencies. It is the responsibility of the user to ensure that the probe is properly calibrated and that an accurate record is kept with each calibration.

SCOPE

The Myron L II is to be calibrated prior to use for the day and details of the calibration are to be recorded in calibration log sheet.

Myron L II Calibration

Specific Conductance Calibration

- Push the COND button
- With KCL 7000 solution, fill the conductivity cell cup completely and dump it 2 times.
- With KCL 7000 solution, fill the conductivity cell cup completely a 3rd time, and let it settle on a reading.
- Push the CAL button.
- Use the up or down buttons to make the display read 7000.
- Hit the CAL button, to accept the calibration.
- If the display does not read 7000, repeat steps 4-6 until it does.

Total Dissolved Solids Calibration

- Push the TDS button
- With the 442-3000 solution, fill the conductivity cell cup completely and dump it 2 times.
- With the 442-3000 solution, fill the conductivity cell cup completely a 3rd time and let it settle on a reading.
- Push the CAL button.
- Use the up or down buttons to make the display read 3000.
- Hit the CAL button, to accept the calibration.
- If the display does not read 3000, repeat steps 4-6 until it does.

PH Calibration

- Push the PH button
- Remove the protective rubber cap from the PH sensor well.
- With the PH7 solution, fill the PH sensor well and the conductivity cell cup completely and dump it 2 times.
- With the PH7 solution, fill the PH sensor well and the conductivity cell cup completely a 3rd time and let it settle on a reading.
- Push the CAL button.
- Use the up or down buttons to make the display read 7.0.
- Hit the CAL button, to accept the calibration.
- If the display does not read 7.0, repeat steps 5-7 until it does.
- Repeat steps 3-8 with the PH4.0 and the PH10.0 solutions

APPENDIX L

Aqua Troll 600 Calibration procedure